

**“COMPARISION OF ANTI-BACTERIAL EFFICACY OF  
CALCIUM HYDROXIDE, DICLOFENAC WITH CALCIUM  
HYDROXIDE AND IBUPROFEN WITH CALCIUM  
HYDROXIDE AGAINST *ENTEROCOCCUS FAECALIS* IN AN  
ENDODONTIC MODEL-AN *IN VITRO* STUDY”**

**By**

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**Dissertation**

*Submitted to*

**KLE Academy of Higher Education and Research  
In partial fulfillment  
Of the requirements for the degree of**

**MASTER OF DENTAL SURGERY**

**In**

**CONSERVATIVE DENTISTRY AND  
ENDODONTICS  
(BRANCH - IV)**

**Under the guidance of**

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**2018-2021**

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*This dissertation is dedicated  
to*

***ALMIGHTY GOD,***

*My Parents,*

***&***

*My Brother*

## ACKNOWLEDGEMENT

*No endeavour can start, continue and complete without the blessings of the Almighty. At the outset, I thank the almighty for always being by my side by bestowing strength and patience to complete the task entrusted. With the completion of my dissertation, I am overcome with a sense of satisfaction and even more gratitude towards all my mentors and well-wishers whom I wish to acknowledge.*

*I would like to express the deepest appreciation to my honourable teacher and guide, **Dr.Neha Dhaded** M.D.S Reader, Department of Conservative Dentistry and Endodontics, KAHER Vishwanath Katti Institute of Dental Sciences, Belagavi. Her patience, encouragement, and meticulous supervision at every step along with her valuable suggestions for improvement enabled me to complete my dissertation which also helped me tremendously during my postgraduate tenure.*

*I was privileged to have **Dr.Jyoti Nagamoti** M.D., Ph.D., Professor, Department of Microbiology, Jawaharlal Nehru Medical College, KAHER, Belagavi as my co-guide for my dissertation. Her constant support and guidance with the microbiological analysis helped me complete my dissertation well in advance.*

*With deep gratitude, I thank **Dr.Preeti Doddwad** M.D.S., Professor and Head of the Department, Department of Conservative Dentistry and Endodontics, KAHER Vishwanath Katti Institute of Dental Sciences, Belagavi for giving enthusiastic encouragement, consistent support and unequivocal help during my post graduation.*

*I am grateful to **Dr.Alka Kale** M.D.S.,Ph.D, Principal, K.L.E. VK Institute of Dental Sciences, Belagavi for extending her help and co-operation towards the completion of this dissertation.*

*I would like to express my solemn gratitude, respect & special debt of thanks to my beloved teachers **Dr.Sonal Joshi** M.D.S. Professor, **Dr.Anand C Patil** M.D.S., Professor, **Dr.Avinash Patil** M.D.S. Reader, **Dr.Sunita Shivanand** M.D.S Reader, **Dr.Shavina Patil** M.D.S Reader, **Dr.Suresh Shenvi** M.D.S Senior lecturer, **Dr.Sneha Patil** M.D.S Senior Lecturer and **Dr.Aniket Chavan** M.D.S Senior lecture, who selflessly devoted their time to me with patience whenever needed. Their colossal contribution all through these years will never be history.*

*My heartfelt thanks to **Dr.Niraj Godbole** M.D.S Senior lecturer, **Dr.Vaidehi Dhopavkar** M.D.S Senior lecturer, **Dr.Mateen Peerzade** M.D.S Senior lecturer and **Dr.Olivia Banerjee** M.D.S Senior lecturer for their sincere help and support given to me during this dissertation.*

*I would also like to extend my thanks to **Mr.U.B. Bolmal** M.PHARM. Assistant Professor, Department of Pharmaceutics, KAHER College of Pharmacy for helping with the preparation of medicament. I would also like to express my solemn gratitude, respect to **Suneel Dodamani** Ph.D, Dr.Prabhakar Kore Basic Science Research Center, KLE Academy of Higher Education and Research for helping me with bacterial study.*

*My sincere thanks to **Mr.Javali**, the statistician, for his services in carrying out the statistical analysis.*

*I am thankful to **Mr.Bagalkoti**, store Incharge, **Mr.Kedar**, Technician, **Mrs.Geeta**, **Mr.Mallappa**, Materials incharge and all **non-teaching staff** of my Department for their immense help during the course of my study.*

*My sincere thanks to my loving seniors **Dr.Ankita**, **Dr.Akash**, **Dr.Suraj**, **Dr.Pratik** and **Dr.Neha** for having generously shared insight and for their unwavering support.*

*I extend my heartfelt gratitude to my colleagues **Dr.Felbin, Dr.Ashish, Dr.Shrutika and Dr.Chaitra** for their constant support and co-operation .*

*I am thankful to my juniors **Dr.Shefali and Dr.Aishwarya** for always being helpful and proof reading my dissertation.*

*I extend my heartfelt thanks to my junior colleagues **Dr.Amruta, Dr.Greeta, Dr.Aishika, Dr.Bhavana, Dr.Mahima, Dr.Priyanka, Dr.Abhijit, Dr.Rohit** for their help and co-operation.*

*I am thankful to my friends **Dr.Anmol Mittal, Dr.Achal Mittal, Neha Kakkar, Dr.Praveen, Dr.Prashasti, Dr.Uday** for their constant support.*

*I am thankful to **Mr. Anand and Mr. Arun** of Shri Vigneshwara Associates, Belagavi for formatting, printing and binding of my dissertation.*

*I would not have completed this dissertation without the unconditional support of my family, who have always been there for me whenever I needed them, with encouragement and love to empower me all the time. I owe everything of what I am and to my parents, **Mr.P.J.Chacko and Mrs.Jessy Chacko**, my younger brother **Mr.Vishal Jose Chacko**, who gave form to all my dreams and aspirations. I would also like to thank my fiance **Dr.Christy George Palathra** for always keeping my morale afloat.*

*I owe every success and joy to them and I humbly acknowledge that everything I am today is because of their love and support.*

Thank you, One and all.

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## ABSTRACT

**Aim and Objectives:** The purpose of this *In vitro* study was to evaluate and compare the antibacterial efficacy of Calcium hydroxide, Diclofenac with Calcium Hydroxide and Ibuprofen with Calcium Hydroxide against *E. faecalis*.

**Study design:** Sixty extracted human permanent maxillary central incisors which fulfilled the inclusion and exclusion criteria were selected for the study. Decoronation of samples was done at CEJ using a diamond disc to standardize the root length to 14 mm with continuous water coolant. A size 10 K-file was used and working length was established 1mm short of the length where 10 K-file exited the apical foramen. Cleaning and shaping were performed using Protaper Universal (DENTSPLY) nickel-titanium rotary instrument up to F3/F4, depending upon the apical gauging. 2ml of 3% Sodium Hypochlorite (NaOCl) was used as an irrigating solution after each instrument followed by 2 mL of 17% EDTA. The final irrigation was done using 5 ml of distilled water. After drying the canals with size 30 absorbent paper points, all external surfaces were made impermeable with nail varnish, except for coronal access. Teeth were then autoclaved at 121°C for 20 min. The prepared samples were placed in an Eppendorf tube for 14 days to inoculate the samples with the test organism i.e. *E.faecalis*. The first root canal sample S-1 was taken immediately after the inoculation of the specimen.

According to the placement of the medicaments, the samples were divided into three groups (n=20) -

GROUP 1: Calcium Hydroxide.

GROUP 2: Diclofenac with Calcium Hydroxide.

### GROUP 3: Ibuprofen with Calcium Hydroxide.

After one week of the medicament placement, the canals were rinsed and cleaned with passive ultrasonic irrigation along with 10 ml of 17% EDTA, followed by 10 ml of 3% NaOCl and a final rinse with distilled water. Sample(S-2) was collected in the same manner as sample 1 for microbiological analysis.

**Results:** There was a statistically significant difference between groups (Calcium Hydroxide, Diclofenac with Calcium Hydroxide, and Ibuprofen with Calcium Hydroxide) based on the reduction in the log CFU in the post medicament samples ( $p < 0.05$ ) at 5% level of significance.

It was found that Diclofenac with Calcium Hydroxide group was superior to Calcium Hydroxide group and Ibuprofen with Calcium Hydroxide group.

**Conclusion:** Within the limitations of this *In vitro* study, Diclofenac with Calcium Hydroxide has shown better antimicrobial activity as compared to Calcium Hydroxide, and Ibuprofen with Calcium Hydroxide. Thus, Diclofenac with Calcium Hydroxide has shown promising results suggesting that it could be used as an alternative intracanal medicament.

**Keywords:** Antibacterial agents; anti-inflammatory nonantibiotics; Calcium Hydroxide; Diclofenac; Ibuprofen.

## LIST OF ABBREVIATIONS

SR.NO	ABBREVIATIONS	FULL FORM
1	RCT	Root Canal Treatment
2	Ca(OH) <sub>2</sub>	Calcium hydroxide
3	PUI	Passive ultrasonic irrigation
4	<i>et al</i>	Additional persons involved in the same study
5	<i>E. faecalis</i>	<i>Enterococcus faecalis</i>
6	Ni-Ti	Nickel Titanium
7	CEJ	Cemento-Enamel Junction
8	NSAIDs	Nonsteroidal Anti-Inflammatory Drugs
9	° C	Degrees Celsius
10	DW	Distilled Water
11	DMSO	Dimethyl sulfoxide
12	MIC	The mean minimum inhibition concentration
13	US scaler	Ultra-Sonic scaler
14	OSHA	Occupational Safety and Health Administration
15	ANOVA	Analysis of Variance
16	SD	Standard Deviation
17	N	Number of specimens
18	p-value	Probability of obtaining a test statistic at least as extreme as the one that was actually observed
19	WL	Working Length
20	MBC	The mean Minimum Bactericidal concentration
21	NaOCl	Sodium Hypochlorite

<b>22</b>	EDTA	Ethylene Diamine Tetra-acetic Acid
<b>23</b>	<	Less than
<b>24</b>	>	Greater than
<b>25</b>	CFU/ml	Colony-forming units per milliliter
<b>26</b>	mm	Millimetre
<b>27</b>	%	Percentage
<b>28</b>	ATCC	American type culture collection
<b>29</b>	BHI	Brain heart infusion
<b>30</b>	pH	power of hydrogen
<b>31</b>	w/w	Weight/Weight

## **TABLE OF CONTENTS**

<b>SL.NO.</b>	<b>PARTICULARS</b>	<b>PAGE NO.</b>
<b>1</b>	<b>INTRODUCTION</b>	<b>1-3</b>
<b>2</b>	<b>AIM AND OBJECTIVES</b>	<b>4</b>
<b>3</b>	<b>RESEARCH HYPOTHESIS</b>	<b>5</b>
<b>4</b>	<b>REVIEW OF LITERATURE</b>	<b>6-12</b>
<b>5</b>	<b>MATERIALS AND METHOD</b>	<b>13-38</b>
<b>6</b>	<b>RESULTS</b>	<b>39-51</b>
<b>7</b>	<b>DISCUSSION</b>	<b>52-62</b>
<b>8</b>	<b>CONCLUSION</b>	<b>63</b>
<b>9</b>	<b>SUMMARY</b>	<b>64-65</b>
<b>10</b>	<b>BIBLIOGRAPHY</b>	<b>66-79</b>
<b>11</b>	<b>ANNEXURES</b>	<b>80-82</b>

## LIST OF TABLES

TABLE NO.	PARTICULARS	PAGE NO.
<b>1</b>	Depicting MIC and MBC score distribution of all the Groups.	<b>39</b>
<b>2</b>	Depicting summary Statistics of the three Groups( (MIC Score).	<b>39</b>
<b>3</b>	Table showing inhibition of bacteria by the 3 Groups through one way ANOVA.	<b>40</b>
<b>4</b>	Table comparing inhibition of bacteria by Tukeys multiple posthoc test .	<b>40</b>
<b>5</b>	Table Showing summary of comparision among the three groups.	<b>41</b>
<b>6</b>	Table comparing bactericidal activity among the 3 Groups through one way ANOVA.	<b>42</b>
<b>7</b>	Table comparing bactericidal activity among the 3 Groups through Tukeys multiple posthoc test.	<b>42</b>
<b>8</b>	Table comparing before and after treatment CFU count by Kolmogorov Smirnov test.	<b>43</b>
<b>9</b>	Depicting summary Statistics of before and after treatment of CFU counts.	<b>44</b>
<b>10</b>	Table comparing mean before and after treatment CFU count by one way ANOVA.	<b>45</b>
<b>11</b>	Table showing comparision of three Groups using mean CFU Score by Tukeys multiple posthoc test	<b>46</b>
<b>12</b>	Comparison of before and after treatment with mean CFU counts in three groups by Dependent t-test	<b>48</b>

## **LIST OF GRAPHS**

<b>SL.NO.</b>	<b>PARTICULARS</b>	<b>PAGE NO.</b>
<b>1</b>	Comparison of the three groups with mean MIC scores	<b>41</b>
<b>2</b>	Comparison of the three groups with mean MBC scores	<b>43</b>
<b>3</b>	Comparison of three groups with mean before and after treatment CFU count	<b>47</b>
<b>4</b>	Comparison of before and after treatment with a mean CFU count in the three groups	<b>49</b>

## LIST OF FIGURES

FIG NO.	PARTICULARS	PAGE NO.
<b>1</b>	The Minimum Inhibitory Concentration (MIC) and Minimum Bactericidal Concentration (MBC) for Diclofenac with Calcium Hydroxide (Group II) and Ibuprofen with Calcium Hydroxide (Group III) carried out against <i>E. faecalis</i> .	<b>18</b>
<b>2</b>	Sixty extracted human maxillary central incisors teeth.	<b>26</b>
<b>3</b>	Debris and calculus removal with an ultrasonic scaler.	<b>26</b>
<b>4</b>	De-coronation at the level of CEJ.	<b>27</b>
<b>5</b>	Materials used for Microbiological assessments.	<b>28</b>
<b>6</b>	Hand and rotary instruments required for root canal preparation.	<b>28</b>
<b>7</b>	Materials required for medicament preparation and placement.	<b>29</b>
<b>8</b>	Materials required for irrigation and medicament removal.	<b>29</b>
<b>9</b>	Minimum Inhibitory Concentration (MIC) of Calcium Hydroxide (Group I) for <i>E. faecalis</i> .	<b>30</b>
<b>10</b>	Minimum Inhibitory Concentration (MIC) of Diclofenac with Calcium Hydroxide (Group II) for <i>E. faecalis</i> .	<b>30</b>
<b>11</b>	Minimum Inhibitory Concentration (MIC) of Ibuprofen with Calcium Hydroxide (Group III) for <i>E. faecalis</i> .	<b>30</b>
<b>12</b>	Minimum Bactericidal Concentration (MBC) of Calcium Hydroxide (Group I) for <i>E. faecalis</i>	<b>31</b>
<b>13</b>	Minimum Bactericidal Concentration (MBC) of Diclofenac with Calcium Hydroxide (Group II) for <i>E. faecalis</i>	<b>31</b>

<b>14</b>	Minimum Bactericidal Concentration (MBC) of Ibuprofen with Calcium Hydroxide (Group III) for <i>E. faecalis</i>	<b>31</b>
<b>15</b>	Working length determination.	<b>32</b>
<b>16</b>	Biomechanical preparation.	<b>32</b>
<b>17</b>	Irrigation using side vented needles.	<b>33</b>
<b>18</b>	Sample inoculation with <i>E. faecalis</i>	<b>33</b>
<b>19</b>	Pre-treatment <i>E. faecalis</i> colonies in G1	<b>34</b>
<b>20</b>	Pre-treatment <i>E. faecalis</i> colonies in G2	<b>34</b>
<b>21</b>	Pre-treatment <i>E. faecalis</i> colonies in G3	<b>34</b>
<b>22</b>	Incubation.	<b>35</b>
<b>23</b>	Mixing of the Medicament	<b>35</b>
<b>24</b>	Placement of medicaments using lentulospiral	<b>36</b>
<b>25</b>	Test samples were kept in incubator for 7 days	<b>36</b>
<b>26</b>	Ultrasonic irrigation to remove the medicament	<b>37</b>
<b>27</b>	Ultrasonic irrigation to remove the medicament	<b>37</b>
<b>28</b>	Post medicament colonies seen in Group 1	<b>38</b>
<b>29</b>	Post medicament colonies seen in Group 2	<b>38</b>
<b>30</b>	Post medicament colonies seen in Group 3	<b>38</b>

## **ANNEXURE**

<b>ANNEXURE NO.</b>	<b>PARTICULARS</b>	<b>PAGE NO.</b>
<b>I</b>	ETHICAL APPROVAL CERTIFICATE	<b>80</b>
<b>II</b>	BIOSTATISTIC CLEARANCE CERTIFICATE	<b>81</b>
<b>III</b>	PLAGIARISM CHECK CERTIFICATE	<b>82</b>

## INTRODUCTION

Bacteria plays a pivotal role in the pathogenesis of apical periodontitis. It leads to pain and tenderness of the affected tooth, which subsequently requires root canal treatment RCT. For optimal treatment outcome complete elimination of bacteria is of utmost importance<sup>1,2</sup>. Cleaning and shaping along with antimicrobial agents lead to the reduction in the bacterial colonies present in the root canal system<sup>3</sup>.

However, studies have shown chemo-mechanical instrumentation alone is unable to abolish the full bacterial colonies in the root canal system<sup>4-7</sup>. To achieve the above mentioned beneficial effect, intracanal medication is advocated which will lead to bacterial reduction<sup>8</sup>.

The gold standard intracanal medicament used during inter-appointment dressing is Calcium Hydroxide due to its therapeutic antibacterial along with its inherent biological properties. Despite having favourable clinical evidence, it has been reported that there is a correlation between Ca(OH)<sub>2</sub> placement with that of increased incidence of tooth fracture, principally reported in teeth with an open apex. Studies have even shown that Ca(OH)<sub>2</sub> is ineffective in recurrent and chronic infection cases<sup>10,11</sup>.

The most habitually encountered bacteria in recurrent and chronic infection cases is *E.faecalis*. Various studies have shown their colonization inside the root canal system as isolated infections. This kind of colonization is most commonly seen in failed RCT cases. It has also been reported that they can infiltrate dentinal tubules to a deep magnitude with a predisposition to form biofilms. This enables it to escape endodontic instrumentation and irrigation. It also renders it impervious to

Ca(OH)<sub>2</sub> due to its potential to combat pH values on the higher side. *E.faecalis* possesses the ability to increase its virulence by inheriting traits such as antibiotic resistance by plasmid transfer <sup>12</sup>.

As stated, the thorough elimination of bacteria is essential for a favourable prognosis following an RCT<sup>13-15</sup>. Studies have shown, that with a qPCR-negative result, healing is seen in up to 79% of RCT cases <sup>15</sup>. Contrary to this, canals which were qPCR-positive exhibited a decreased healing rate of 45% <sup>15</sup>. Hence, there is a need to improve the efficacy of Ca(OH)<sub>2</sub>. To attain improvement in the anti-bacterial effect of Ca(OH)<sub>2</sub>, additions such as NSAIDs and antibiotics have been advocated for a favourable treatment outcome <sup>16</sup>.

The conventionally used class of analgesics NSAIDs– may possess additional therapeutic properties such as anti-bacterial efficacy through inhibition of bacterial DNA synthesis or impairment of membrane activity <sup>17-20</sup>; prevention of bacterial colonization and biofilm formation by interfering with quorum sensing of bacteria <sup>21</sup>; and anti-plasmid activity<sup>22</sup>. This finding is merely one in the long line of research investigating the antibacterial effects of non-antibiotic agents, commencing with Ehrlich's (1854–1915) discovery of the antibacterial effects of the Phenothiazine compound, methylene blue dye. The term non-antibiotics was coined by Kristiansen and Amaral (1997) to refer to drugs developed to treat noninfectious diseases but have shown to exhibit anti-microbial activity <sup>23</sup>.

Other anti-inflammatory agents such as Corticosteroids (Ledermix) are already in use as inter-appointment intracanal medicaments. They have shown efficacy in reducing post-operative pain compared to Ca(OH)<sub>2</sub> <sup>24</sup>, but they may compromise host immune response <sup>25</sup>. Further studies have shown that long term exposure of the

aqueous tetracycline component present in Ledermix causes enamel discolouration <sup>26</sup>.

It has been established that 1-25 hrs of enamel exposure to this aqueous tetracycline component in long treatment protocols reduces the microhardness of enamel <sup>26</sup>.

Till present there is no single antibiotic that is effective against all types of microorganisms present within the diseased tooth, therefore local antibiotics are not routinely recommended <sup>27</sup>. Studies have reported that routinely used antibiotics cause teeth discoloration due to the presence of minocycline, making it un-esthetic when it is used in the treatment of anterior teeth<sup>28, 29</sup>. Furthermore, it is also seen that in regenerative endodontics and non-surgical management of large periapical lesions, the longterm use of triple antibiotic paste causes a negative impact on radicular dentin by increased demineralization and reduction in microhardness of the dentin <sup>26,30</sup>.

Therefore, incorporating commonly prescribed NSAIDs such as Diclofenac or Ibuprofen as a component in inter-appointment dressing has the potential to utilize beneficial effects of NSAIDs i.e anti-inflammatory action, local analgesia, and potential anti-bacterial action, and also to improve on the properties of Ca(OH)<sub>2</sub> (limited anti-bacterial action).

There are very limited studies done which have evaluated the anti-bacterial efficacy of NSAIDs with Ca(OH)<sub>2</sub> against *E.faecalis*. Considering all the above-mentioned concepts, this study was formulated to evaluate and compare the antibacterial efficacy of NSAIDs with Ca(OH)<sub>2</sub> against *E.Faecalis*.

## **AIM AND OBJECTIVES**

### **AIM**

To evaluate and compare the antibacterial efficacy of three different intra-canal medicaments Calcium hydroxide, Diclofenac with Calcium Hydroxide and Ibuprofen with Calcium Hydroxide against *E. faecalis*.

### **OBJECTIVES**

- To determine the minimum bactericidal concentration (MBC) of Diclofenac with Calcium Hydroxide and Ibuprofen with Calcium Hydroxide.
- To assess the antibacterial activity of Calcium Hydroxide, Diclofenac with Calcium Hydroxide and Ibuprofen with Calcium Hydroxide against *E. faecalis* using semi-quantitative analysis.
- To compare the antibacterial activity of Calcium Hydroxide, Diclofenac with Calcium Hydroxide and Ibuprofen with Calcium Hydroxide as an intra-canal medicament against *E. faecalis*.

## **HYPOTHESIS**

### **NULL HYPOTHESIS: -**

There is **no difference** in the antibacterial efficacy of Calcium Hydroxide, Diclofenac in combination with Calcium hydroxide and Ibuprofen in combination with Calcium Hydroxide when used as an intra-canal medicament.

### **ALTERNATE HYPOTHESIS: -**

There is a **difference** in the antibacterial efficacy of Calcium Hydroxide, Diclofenac in combination with Calcium Hydroxide and Ibuprofen in combination with Calcium Hydroxide when used as an intra-canal medicament.

## REVIEW OF LITERATURE

1. A clinical review on the current concepts of root canal infection and the consequences for endodontic treatment has emphasized the usage of intracanal medicament in cases of apical periodontitis. The review emphasizes the various fruitful and favourable properties of the medicament that renders the root canal system bacteria-free resulting in better treatment outcome<sup>32</sup>.
2. A study was done to evaluate the antibacterial effect of calcium hydroxide paste in distilled water as an intracanal medicament in twelve cases. It was found that 54.5 % of the cases became asymptomatic and sterile in 4 visits, 27.2 % in 7 visits, while 18.20 % were failures. It was concluded that calcium hydroxide could be successfully used as an intracanal medicament in highly infected canals<sup>33</sup>.
3. Antibacterial activity of 2 formulations of calcium hydroxide pastes (Calen™ and Calen with p-chlorophenol- PMCC-Calén™) on *E.faecalis* grown in contact with dentinal walls of root canal systems was evaluated. The results of the study concluded that intracanal usage of Calcium Hydroxide pastes after chemo-mechanical preparation of root canals, were capable of eliminating *E. faecalis*, which is the most resistant bacteria known for re-treatment cases<sup>34</sup>.
4. An *In vivo* study was done to evaluate the antibacterial efficacy of Calcium Hydroxide when used as an intracanal medicament for a short period of time. The study showed that a 10-minute application of the dressing was ineffective, when the same was applied for seven-days, it was able to eliminate the bacteria that has survived the biomechanical preparation of the canal. After chemomechanical preparation of the root canals, the presence of bacteria was

seen in 9 out of the 18 canals which were dressed with Calcium Hydroxide for a period of seven days and in 6 of the 12 canals treated with Calcium Hydroxide for around 10 minutes. No bacteria were found in tests following the removal of the dressing among the 18 canals that had been dressed for one week, neither from samples which had been sealed without an antibacterial dressing for an extended week(1-5 weeks). Application of Calcium Hydroxide for 10 minutes was ineffective in 6 root canals since bacteria persisted at the second appointment <sup>35</sup>.

5. An *In-vitro* study was done to check the long term effect of Calcium Hydroxide dressing. The review tells us about the various beneficial antibacterial properties of  $\text{Ca(OH)}_2$  when used in root canal treatment as an intracanal medicament. Furthermore, it explains the reason for the antimicrobial effect of  $\text{Ca(OH)}_2$ , which is due to the release of hydroxyl ions when it comes in contact with aqueous fluids. Even though  $\text{Ca(OH)}_2$  shows effectiveness against a wide range of endodontic pathogens, but it's efficacy against *Enterococcus faecalis* and *Candida albicansis* effectively less. The studies end with the conclusion that the addition of vehicles or other agents might contribute to an increase in the antimicrobial effect of  $\text{Ca(OH)}_2$  <sup>36</sup>.
6. A meta-analysis study was done in patients undergoing root canal treatment for apical periodontitis, to determine the antibacterial activity of Calcium Hydroxide when used as an intracanal medication. Eight studies were chosen and added in the review, covering 257 cases with a sample size of 18 to 60 cases. A statistically significant difference between pre and the post-medicated canal was seen among six studies. The study concluded that Calcium

Hydroxide when assessed by culture techniques, has limited efficacy in bacteria eradication from the root canal <sup>37</sup>.

7. An *In-vitro* study was done which stated that the teeth treated with Calcium Hydroxide as an intracanal medicament were more susceptible to fracture. Edge chipping was the test of choice used to measure the resistance of brittle materials to fracture. Chip resistance of the teeth may depend on both the factors i.e. fracture resistance and the hardness of dentin. The study uses an edge chipping test to demonstrate the fracture resistance of human dentin exposed to calcium hydroxide for up to 60 days. Twelve teeth that were recently extracted were divided into three experimental groups and a control group with varying calcium hydroxide exposures. All the teeth used in this study underwent pulpectomy via a standard protocol. The study expected the Ca(OH)<sub>2</sub> treated teeth to be more susceptible to fracture in comparison to the control group but the results showed that the edge chip resistance was improved in all Calcium Hydroxide exposed test groups. The reason for the increase could be due to change in the molecular structure of the dentin which increased fracture resistance of the dentin. Another possible explanation could be that increase in chip resistance in the earlier study might be due to other forms of a mechanism other than chipping <sup>38</sup>.
  
8. An *In-vitro* examination was done among Diclofenac, Ibuprofen, Calcium Hydroxide and Amoxicillin to evaluate their antibacterial effect against *E. faecalis*. The Agar diffusion test and tube dilution method were the tests of choice to evaluate the antibacterial activity of materials. Mixtures of 400 mg/ml of materials were prepared. 10 Muller-Hinton agar culture plates were

used onto which the bacteria were seeded. In each well punched in agar plates, 30 microliters of each test material were placed. Once incubation was done, the zone of bacterial inhibition was measured. The Agar dilution method was used to find Minimum inhibitory concentration (MIC) of the test materials. The results showed the greatest antibacterial activity by antibiotics (Amoxicillin, Gentamycin) followed by NSAIDs (Ibuprofen, Diclofenac) and  $\text{Ca(OH)}_2$  failed to show any antibacterial activity. Distinct antibacterial activity was shown by Diclofenac and Ibuprofen against *E. faecalis* in concentrations of 50  $\mu\text{g/ml}$  and above<sup>39</sup>.

9. Another study was done in an endodontic model to investigate and compare the anti-bacterial effectiveness of intracanal medicaments Ibuprofen, Diclofenac, and Calcium Hydroxide against *E. faecalis*. The study included a total of 76 single-rooted mandibular premolar teeth which were decoronated and instrumented up to F4-ProTaper rotary. The roots were subjected to autoclave (121°C for 20 min), placed in Eppendorf tubes, and contaminated with *E. faecalis* for 14 days. Paper point sampling was used to record the Colony-forming unit (CFU) counts before (CFU-1), and after intracanal medication (CFU-2). Distilled water (1:1 w/v) was mixed with Group-1: Ibuprofen, Group-2: Diclofenac, Group-3:  $\text{Ca(OH)}_2$ , which was then placed into root canals, temporarily sealed and incubated (37°C; 7 days). Group-4 being the control group, received no medicament. The results demonstrated a greater antibacterial effect against *E. faecalis* by the anti-inflammatory nonantibiotics (Ibuprofen, Diclofenac)<sup>40</sup>.

- 10.** An *In-vitro* study was done to evaluate and compare the different combination of NSAIDs and whether they altered the pH of Calcium Hydroxide. The groups analyzed were group 1: Calcium Hydroxide paste with Propylene Glycol, group 2: Calcium Hydroxide paste with Propylene Glycol + 5% Diclofenac Sodium, group 3: Calcium Hydroxide paste with Propylene Glycol + 5% Ibuprofen, group 4: Calcium Hydroxide paste with Propylene Glycol + 5% Ciprofloxacin and group 5: positive control (without medication) The pH was measured with a calibrated pH meter, at time intervals of 3, 24, 72, and 168 hours. The biofilm was induced in 30 bovine dentin blocks for 21 days, for microbial analysis. The pastes were placed on the blocks with biofilm for 7 days. The specimens were analyzed with a laser scanning confocal microscope. Results demonstrated statistically significant differences ( $P < .05$ ) in comparison with the positive control <sup>41</sup>.
  
- 11.** A review article stated that the process of the formulation was greatly influenced by the solubility properties of drugs. The problem of solubility could be solved by different approaches. To improve the solubility of poorly water-soluble drugs, the complexation technique had been employed. Cyclodextrin can interact with appropriate size drug molecules which lead to the formulation of inclusion complexes. To determine the rightful utilization of cyclodextrins as complexing, solubility enhancing agents, a comprehensive literature survey was made. This review discusses the various complexation techniques and highlights the applications with these approaches <sup>42</sup>.

12. An *In-vivo* study was done on asymptomatic apical periodontitis cases to evaluate and compare the effectiveness of Ca(OH)<sub>2</sub> paste against microorganisms when combined with Ibuprofen and Ciprofloxacin. The study comprises of 45 patients who were subdivided into three groups on the basis: Ca(OH)<sub>2</sub>: 1 gm Ca(OH)<sub>2</sub> powder with 1 mL propylene glycol, Ca(OH)<sub>2</sub> + Ibuprofen: 50 mg of ibuprofen was added into 950 mg Ca(OH)<sub>2</sub> powder and mixed with 1 mL propylene glycol, Ca(OH)<sub>2</sub> + Ciprofloxacin: 50 mg of Ciprofloxacin was added into 950 mg Ca(OH)<sub>2</sub> powder and mixed with 1 mL propylene glycol. The microbiological samples were taken before (S1) and after chemo-mechanical procedures (S2). The intracanal medicaments were placed using K-file after canals were prepared. The second visit of patients was scheduled after 7 days post-medicament wherein the medicament was removed mechanically and the samples (S3) were collected. The bacterial counting was done using a quantitative real-time polymerase chain reaction. The quantitative reduction from S1 to S3 and from S2 to S3 of Ca(OH)<sub>2</sub> + Ciprofloxacin was superior when compared to other groups ( $P < 0.05$ )<sup>43</sup>.
13. A Systematic review was done to study *Enterococcus faecalis*, which is the most common bacteria seen in re-treatment cases. Two databases, PubMed and Google Scholar were searched using specific inclusion and exclusion criteria in this Systemic review. Among 2943 studies that were found, only 11 met the inclusion criteria, and these 11 were included in the review for further analysis. The 11 studies showed a prominent distribution of *Enterococcus faecalis* within the root canal. The study investigated various aspects of *Enterococcus faecalis* and showed its association as the primary pathogen with

endodontic treatment. Moreover, the *Enterococcus faecalis* has the characteristic properties to escape the disinfection means<sup>44</sup>.

14. An *In-vitro* study was done to evaluate and compare the dentinal tubule penetration of Calcium Hydroxide(CH) and triple antibiotic paste(TAP) when performed with distilled water (DW) or a low surface tension liquid (i.e. propylene glycol(PG) ). A total of 40 premolars were taken and were standardized to 14 mm root length. The roots were randomly divided into 4 groups (n = 10) according to the root canal medicaments and the vehicles used: group 1:TAP + DW, group 2: TAP + PG, group 3: CH + DW, and group 4:CH + PG. Using a Lentulo spiral the root canal medicaments were applied into the root canals once they were labeled with 0.1% rhodamine. The specimens were placed into acrylic blocks, after which 1-mm sections were taken from the middle third of the root. The samples were studied under a confocal laser scanning microscope. The results showed that CH had a lower penetration area than TAP regardless of the vehicle used ( $P < .05$ )<sup>45</sup>.

## **MATERIALS AND METHODS**

### **SOURCE OF DATA:**

The study was conducted in the Department of Conservative Dentistry and Endodontics,

V K Institute of Dental Sciences, KLE Academy of Higher Education & Research, Belagavi Karnataka.

Laboratory procedures were carried out in the College of Pharmacy, KAHER Belagavi Karnataka.

The microbiological study was done at:

- Jawaharlal Nehru Medical College, Department of Microbiology, KAHER Belagavi Karnataka.
- Dr.PrabhakarKore Basic Science Research Center, KLE Academy of Higher Education and Research, Belagavi.

### **INCLUSION CRITERIA:**

- Extracted human single-rooted single canal maxillary anterior teeth with patent canals.
- Teeth with apical width corresponding to #20 K-file or less.
- Teeth with straight canals.

**EXCLUSION CRITERIA:**

- Teeth with Carious lesions.
- Teeth with apical width more than #20 K-file size.
- Teeth with calcified canals.
- Teeth with fracture/crack or a restoration.
- Teeth with anatomic variations.

**ARMAMENTARIUM USED FOR THE STUDY**

The following armamentarium was used in the study. The armamentarium used was divided into the following steps as follows:

[A] Armamentarium for MIC, MBC, and microbiological processing

- Diclofenac powder(College of Pharmacy, KAHER Belagavi)
- Ibuprofen powder(College of Pharmacy, KAHER Belagavi)
- ATCC strains of *Enterococcus faecalis* ((ATCC 29212)
- Weighing scale (Citizen CY220, Citizen Scale(I) Pvt. Ltd., Malad, Mumbai)
- Eppendorf tubes (Tarsons Products Pvt. Ltd., West Bengal)
- Pipette with micro pipette tips (Tarsons Products Pvt. Ltd., West Bengal)
- Brain heart Infusion (BHI) Broth(HiMedia laboratories Pvt Limited, Mumbai)
- Incubator (Yorco, York Scientific Industries, India)
- Blood Agar plates
- Platinum Inoculum loops
- Electric loop sterilizer (HiMedia laboratories Pvt Limited, Mumbai)
- Thioglycolate Broth ((HiMedia laboratories Pvt Limited, Mumbai)

[B] ARMAMENTARIUM FOR COMBINATION PREPARATION

- Beaker (Borosil Glass Works Limited, Mumbai)
- Whatman Qualitative filter paper (Sigma- Aldrich Co., USA)
- Petri dish
- Spatula
- Weighing scale (Citizen CY220, Citizen Scale(I) Pvt. Ltd., Malad, Mumbai.
- Diclofenac powder (College of Pharmacy, KAHER Belagavi)
- Calcium Hydroxide (College of Pharmacy, KAHER Belagavi)
- Ibuprofen powder (College of Pharmacy, KAHER Belagavi)
- Cyclodextrin (College of Pharmacy, KAHER Belagavi)
- Ethanol (College of Pharmacy, KAHER Belagavi)
- Distilled water (BenzerMultitech India Private Limited, Pune, India)

[C] Armamentarium for *In-vitro* study

- Human permanent maxillary central incisors teeth (n=60)
- 0.1% Thymol solution (S D FINE-CHEMICALS LIMITED, MUMBAI)
- 3% Sodium hypochlorite (VISHAL DENTOCARE, AHMEDABAD)
- Distilled water (NICE LIF Intracanal medicament- Diclofenac with Calcium Hydroxide (1mg) Calcium Hydroxide (1mg) Ibuprofen with Calcium Hydroxide(1mg)
- 17% EDTA (CANALARGE,AMMDENT PUNJAB)
- Paper points (DIADENT GROUP INTERNATIONAL, KOREA)
- Cavit G - (3M, ESPE GERMANY)
- Endoaccess bur (DENTSPLY, SWITZERLAND)
- 20 K file (MANII INC, JAPAN)

- Protaper Universal nickel-titanium files (DENTSPLY MAILLEFER, SWITZERLAND)
- Airotor (NSK, JAPAN)and Endomotor (DENTSPLY X-SMART)
- Micromotor (NSK, JAPAN)
- Lentulospiral (No-25) (MANI INC, JAPAN)
- 5ml 27-gauge syringe (DISPOVAN, HINDUSTAN SYRINGES LTD FARIDABAD)
- Diamond disc
- Autoclaved (Confident Dental EquipmentsPvt. Ltd., India)
- Pluggers (Sybron endo)
- Tweezer
- Cement spatula
- Contra angle micromotorhandpiece (NSK, Japan)
- RC Prep (Premier Dental)

**METHODOLOGY:**

To get an effective and validated concentration of the medicament Minimum inhibitory concentration and Minimum bactericidal concentration test were done against *Enterococcus faecalis*.

A) DETERMINATION OF MINIMUM INHIBITORY CONCENTRATION -

BROTH DILUTION METHOD: <sup>72</sup>

- Minimum Inhibitory Concentration (MIC) was assessed against the ATCC 29212 strain of *Enterococcus faecalis*.
- Inoculum of standard strains of organisms was prepared as per 0.5 McFarland Standard.

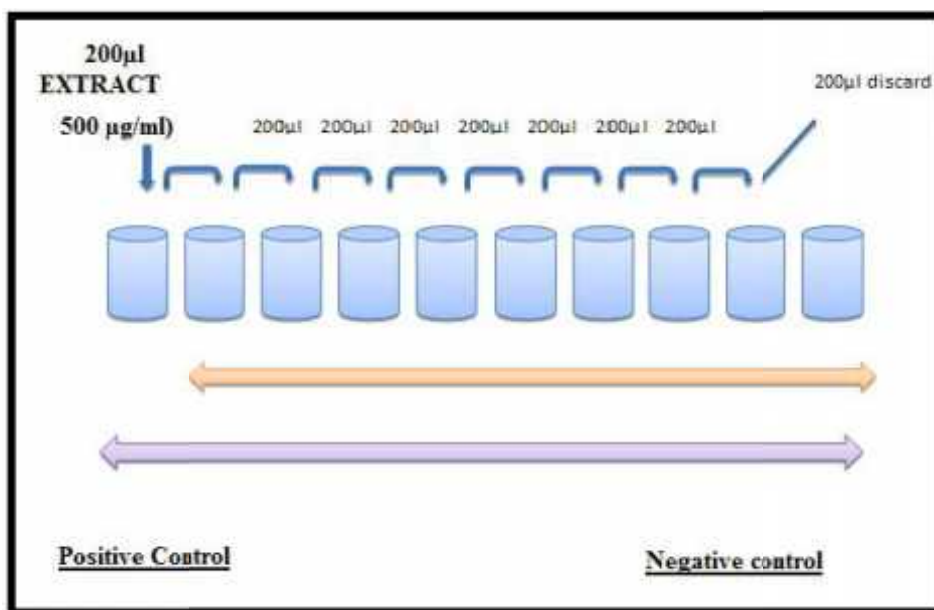
**Methodology:**

Diclofenac and Ibuprofen were taken in combination with Calcium Hydroxide and weighed. These drugs were dissolved in 0.5 ml of Dimethyl sulfoxide(DMSO). Ten graduated micro-centrifuge tubes were taken for the study after being sterilized. These microcentrifuge tubes were designated with numbers from 1 to 10. All the autoclaved centrifuge tubes were arranged systematically following their designated number.

Vertical laminar flow was used to follow the Standard Operating Protocols. Minimum inhibitory Concentration procedure was carried out in the Vertical Laminar flow. For the MIC test, 200 $\mu$ L of plain Brain Heart Infusion (BHI) broth was added to all the 10 tubes with the help of micropipette, this was followed by the addition of 200 $\mu$ L of respective test medicament to the 1st tube. The broth and the test medicament were thoroughly mixed using the micropipette. From the 1st tube 200 $\mu$ L was pipetted and added to the 2nd tube. Again from the 2nd tube 200 $\mu$ L was pipetted

and added to the 3rd tube. This tube dilution method was repeated until the tube 9<sup>th</sup> tube. For the 9th tube, 200µL was pipetted and discarded. Here, the 1st tube is acting as a positive control and the 10th tube is a negative control. Finally, 100µL of inoculum of *E. faecalis* (culture) was added to all the 10 tubes.

All the graduated microcentrifuge tubes were incubated in a CO<sub>2</sub> jar for 48 hours. The MIC was taken as the lowest concentration that prevented the growth of the bacteria for both the groups. To confirm the inhibitory concentration, each of these serial dilutions was plated on blood agar culture plates under laminar airflow for both Group II (Diclofenac with Ca(OH)<sub>2</sub> group) and Group III (Ibuprofen with Ca(OH)<sub>2</sub> group). The culture plates were incubated for 24 hours. Colony-forming units were recorded after the respective times of incubation with the help of Eliza Reader along with manual counting of the colonies.



**Fig.1 The Minimum Inhibitory Concentration (MIC) and Minimum Bactericidal concentration (MBC) for Diclofenac with Calcium Hydroxide (Group 2) and Ibuprofen with Calcium Hydroxide (Group 3) carried out against *E. faecalis***

**MIC Results**

<i>E.faecalis</i>	200µl	100	50	25	12.5	6.25	3.125	1.56	0.78	0.39
D+CH	S	S	S	R	R	R	R	R	R	R
I+CH	S	S	R	R	R	R	R	R	R	R

- S-Sensitive R-Resistant
- Sensitive- the organism is inhibited by the concentration of the drug that is achieved using the usual dosage.
- Resistant- the organisms are resistant to the usually achievable serum drug levels
- MIC was noted by the visual turbidity of the tubes both before and after incubation.
- MIC is the lowest concentration which prevents visible growth of the organism.
- Sensitive implies that the organism is inhibited by the concentration of the drug that is achieved using the usual dosage.

**MBC Results**

<i>E.faecalis</i>	200µg/ml	100	50	25	12.5	6.5	3.125	1.56	0.78	0.39
D+CH	00	00	00	10	30	50	80	120	150	200
I+CH	02	02	03	20	20	30	40	50	80	150

- MBC was noted by the absence of bacterial growth in agar plates both before and after incubation.

**PREPARATION OF MEDICAMENT**

The medicament was prepared in the Department of Pharmaceutics, College of Pharmacy, KLE Academy of Higher Education and Research, Belagavi

**COMPLEXATION METHOD**

The intracanal medicament that was taken for this study was not water-soluble, Therefore the complexation method was chosen to make the medicament's more water-soluble. <sup>42</sup>Complexation is an extensively used technique in the pharmaceutical field to improve the solubility of several pharmaceutical ingredients, to increase the bioavailability of poorly water-soluble drugs. Cyclodextrin is used to increase the solubility of the water-insoluble drug through inclusion complexes formulation. The mentioned compound has a hydrophobic cavity that produces inclusion complexes by trapping a variety of molecules within it. The advantages of drug complexes with cyclodextrin are increased solubility, enhanced bioavailability, improved stability, masking of bad taste or odour, reduced volatility reduced side effect, the possibility of a drug release system. In the present study, test medicaments

were mixed with cyclodextrin, which is an (1,4)-linked oligosaccharides of  $\alpha$ -D-glucopyranose and organic solvent. The obtained mixture was kept in a hot air oven for 2 hours for drying. The dried samples were scrapped and collected for use.

For the *In-vitro* study Diclofenac with  $\text{Ca(OH)}_2$  and Ibuprofen with  $\text{Ca(OH)}_2$  ratio was decided based on the MIC and MBC values of the combination of Diclofenac and Calcium Hydroxide, Ibuprofen and Calcium Hydroxide.

The Diclofenac with  $\text{Ca(OH)}_2$  was prepared in 33.3 %w/w.

The Ibuprofen with  $\text{Ca(OH)}_2$  was prepared in 58.3 w/w.

$\text{Ca(OH)}_2$  was prepared in 72.2% w/w.

#### **Preparation of bacterial sample: Test Microorganism**

- The facultative strain of *Enterococcus faecalis* (ATCC 29212) were grown on brain heart infusion agar plate (with 5% defibrinated sheep blood). Microorganisms were subcultured on nutrient agar medium to confirm their purity. The facultative strain of *Enterococcus faecalis* was inoculated individually into a tube containing 5ml of sterile 85% saline. The suspension was adjusted spectrophotometrically at 800nm to match the transmittance of 90T (equivalent to 0.5 McFarland scale =  $1.5 \times 10^8$  C.F.U).

## **METHODOLOGY**

- Sixty permanent human single-rooted maxillary central incisors were chosen and maintained conferring to OSHA (Occupational Safety and Health Administration) guidelines. Calculus and soft tissue debris were removed with an ultrasonic scaler and 0.1% Thymol(S D FINE-CHEMICALS LIMITED, MUMBAI) was utilized as storage solution until use.

- **SAMPLE PREPARATION**

Decoration of samples was done at CEJ using a diamond disc to standardize the root length to 14 mm with continuous water coolant. A size 10 K file was used and working length was established 1mm short of the length where 10 K file exited the apical foramen. Cleaning and shaping were performed using Protaper Universal (DENTSPLY) nickel-titanium rotary instrument up to F3/F4, depending upon the apical gauging. 2ml of 3% Sodium Hypochlorite (NaOCl) was irrigated after each instrument followed by irrigation with 2 mL of 17% EDTA and was allowed to remain for 1 min. The final irrigation was done using 5 ml of distilled water. After drying with size 30 absorbent paper points, All external surfaces were made impermeable with nail varnish, except for coronal access. Teeth were then autoclaved 121°C for 20 min. The prepared samples were placed in an Eppendorf tube for 14 days to inoculate the samples with the test organism i.e.*E.faecalis*.

### **Collection of Samples**

The first root canal sample S-1 was taken immediately after the inoculation of the specimen. The procedure that was followed -Glucose broth was injected into the root canal and a size 10 or 15 K- H file was pumped circumferentially to 1mm short of the estimated working length. Test tubes containing samples were incubated at

37<sup>0</sup>C for 30 minutes and shaken vigorously in a vortex mixer for 60 seconds. Before inoculation samples were subjected to serial ten-fold dilution in Brain heart infusion broth. 10μL of the sample was inoculated on blood agar and incubated aerobic chamber for 24 hours. The colony-forming units were counted.

The number of colonies was counted by semi quantitation <sup>31</sup>. The colony count was represented as

$$\text{No. of colonies} \times 1000 = \text{Colony forming units (CFU)/ml}$$

For the intracanal medicament placed, 60 specimens were divided randomly divided into groupsof 20 each

Group 1- Ca(OH)<sub>2</sub>

Group 2-Ca(OH)<sub>2</sub> + Diclofenac

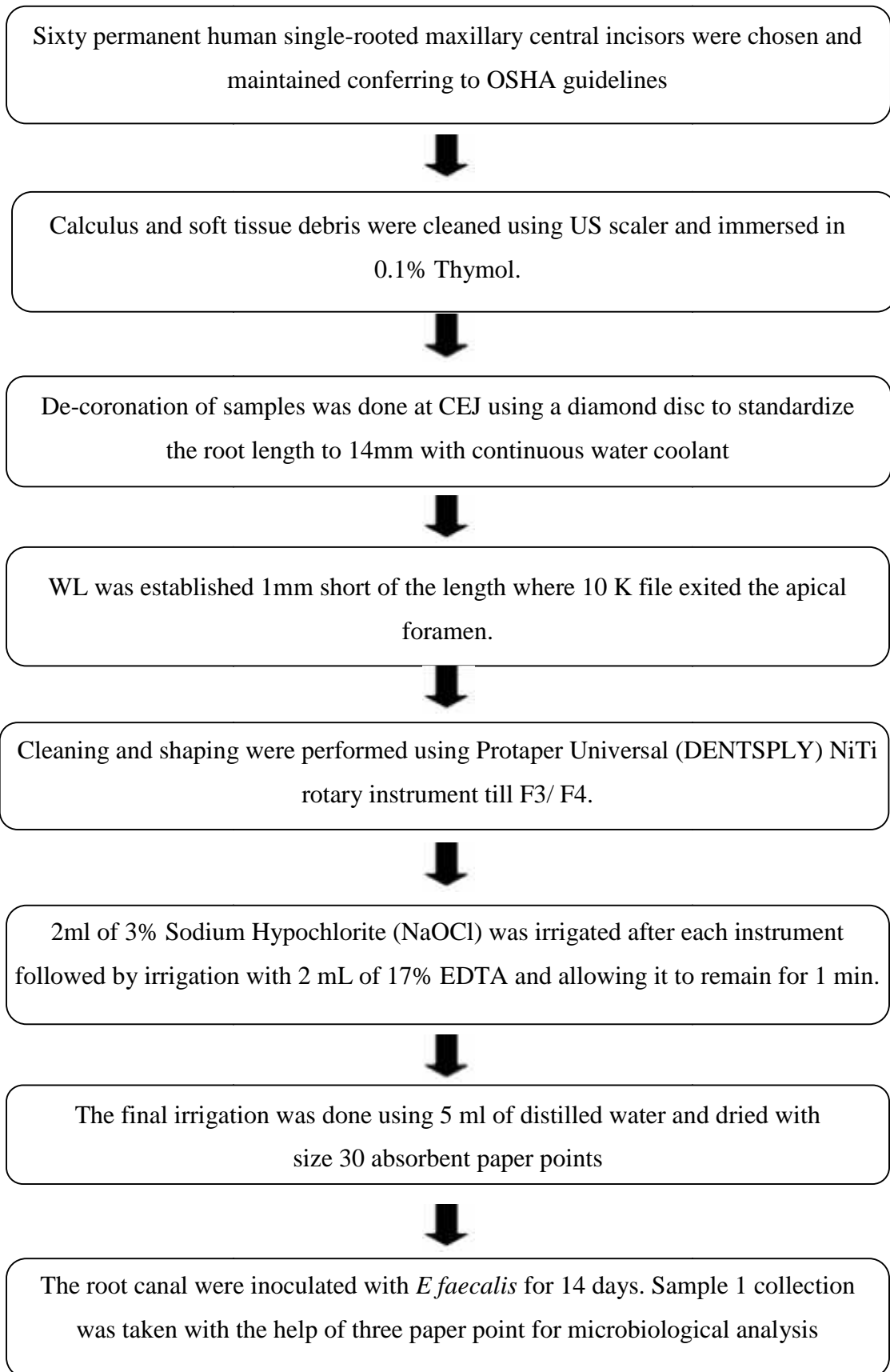
Group3- Ca(OH)<sub>2</sub> + Ibuprofen

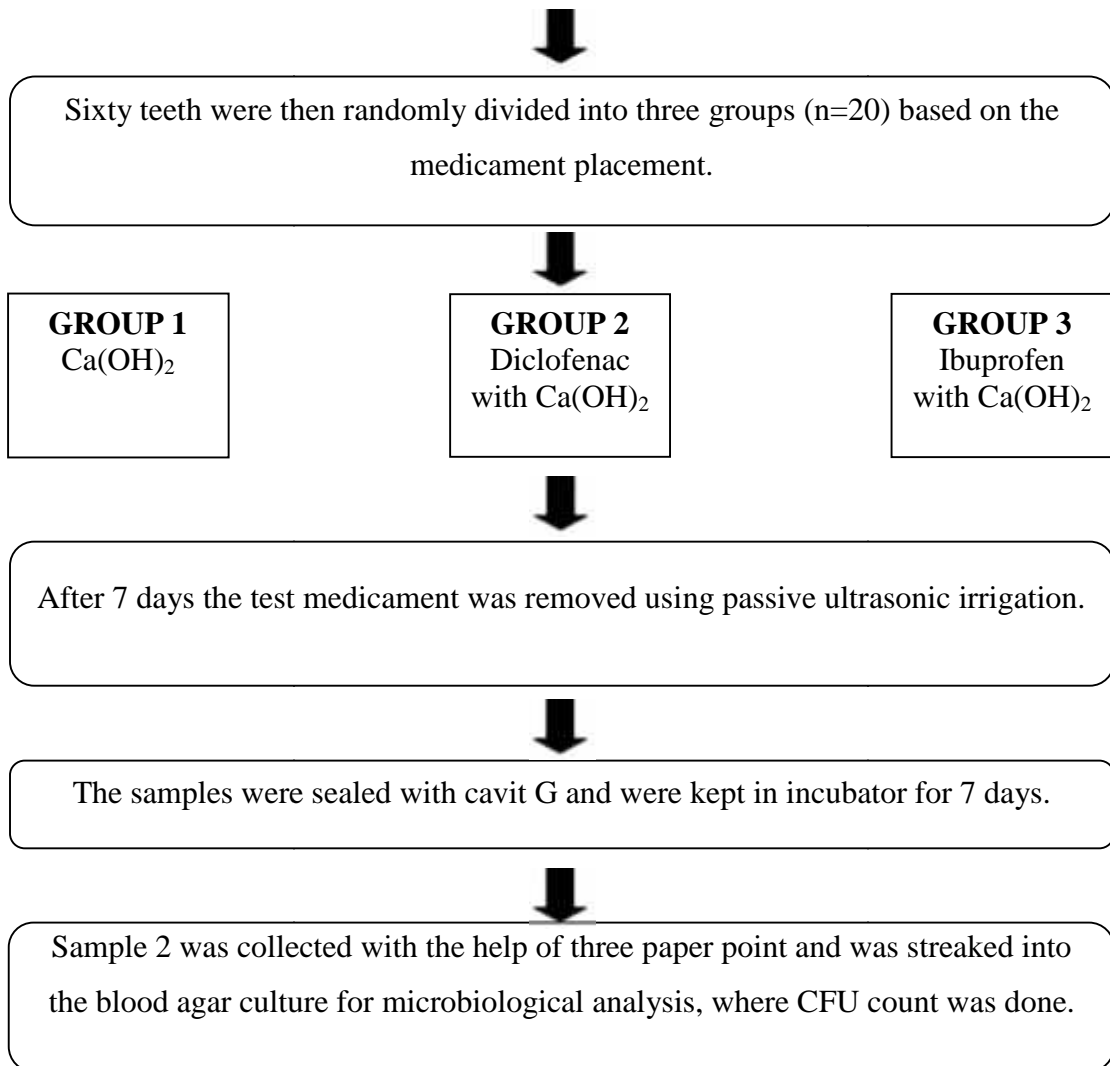
### **Mixing of Test Medicaments**

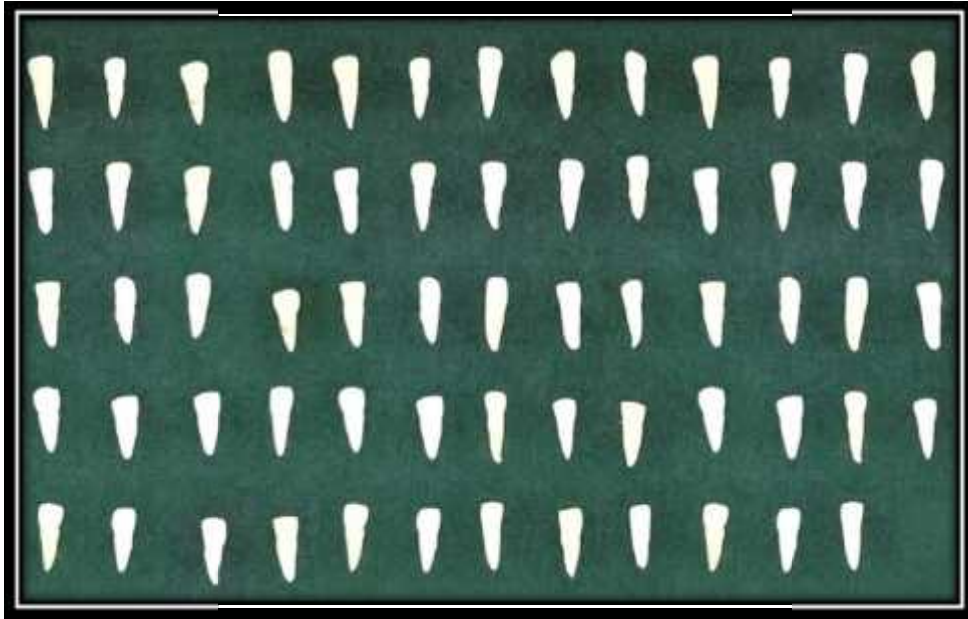
The medicament powder obtained from the pharmacy was mixed with saline in a 1:1 ratio.The prepared pastes of the medicament were carried into the canal using lentulospiral (Mani Inc, Tachigi-ken, Japan) and condensed with hand pluggers (Sybron endo). Cavit G was placed to seal the specimen. The specimens were stored in a humidifier at 37<sup>0</sup>C in 100% humidity for 7 days to simulate clinical conditions. After one week, the canals were rinsed with 10 ml of 17% EDTA followed by 10 ml of 3% NaOCl and final irrigation with distilled water and passive ultrasonic irrigation.

Sample(S-2) was collected in the same manner as sample 1 for microbiological analysis <sup>31</sup>.

**FLOWCHART DEPICTING THE STUDY DESIGN**







**Fig 2:**Total Sample Size (n=60)



**Fig 3:** Removal of debris and calculus using ultrasonic scaler



**Fig 4:** Decoronation of sample at the level of CEJ

**MATERIALS AND ARMAMENTARIUM**



**Fig 5:**Materials used for microbiological assessment



**Fig 6:**Hand and rotary instruments required for root canal preparation



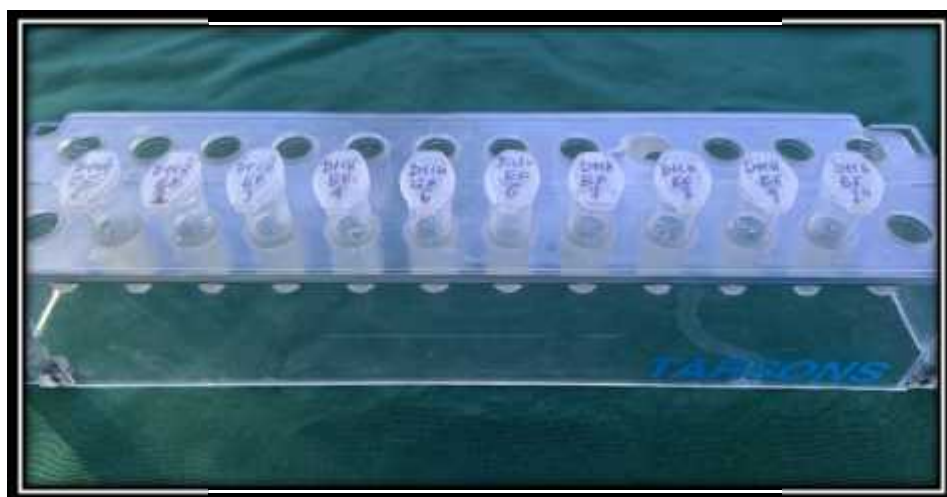
Fig 7: Materials required for medicament preparation and placement



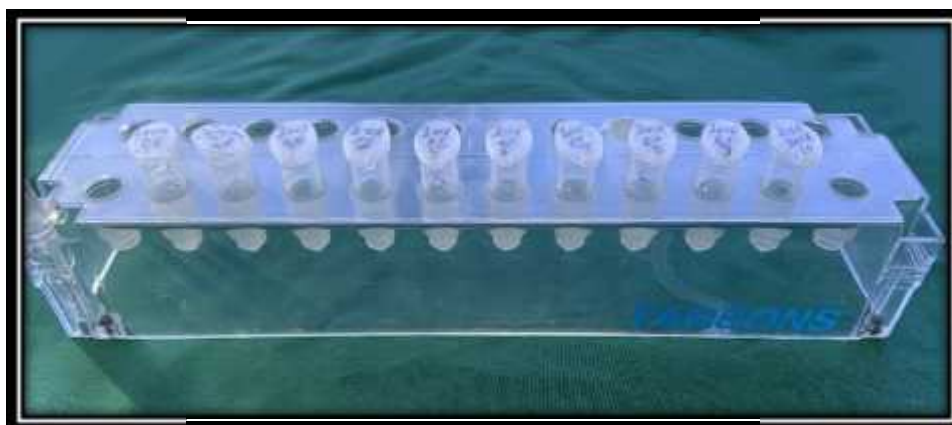
Fig 8: Materials required for irrigation and medicament removal



**Fig 9:** MIC of group 1



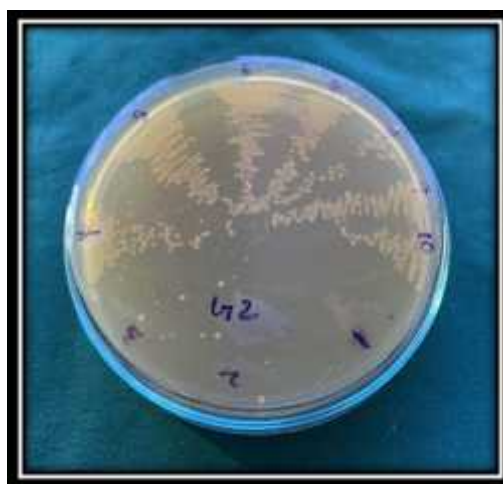
**Fig 10:** MIC of group 2



**Fig 11:** MIC of group 3



**Fig 12:** MBC of group 1



**Fig 13:** MBC of group 2



**Fig 14:** MBC of group 3



**Fig 15:** Working length determination



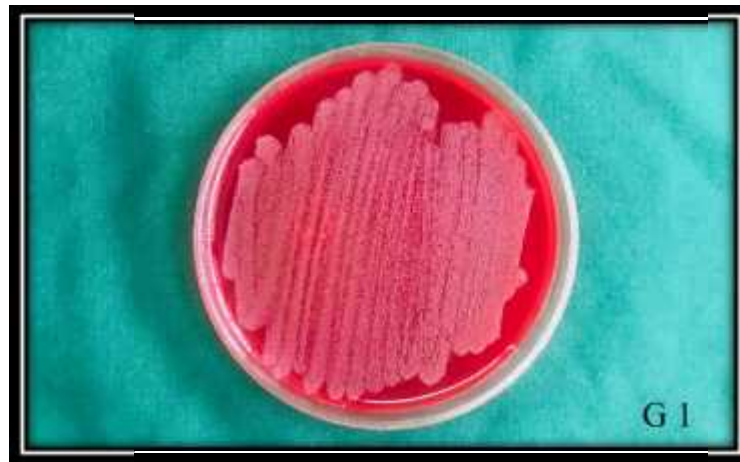
**Fig 16:** BMP using F4



**Fig 17:** Irrigation using 5 ml syringe



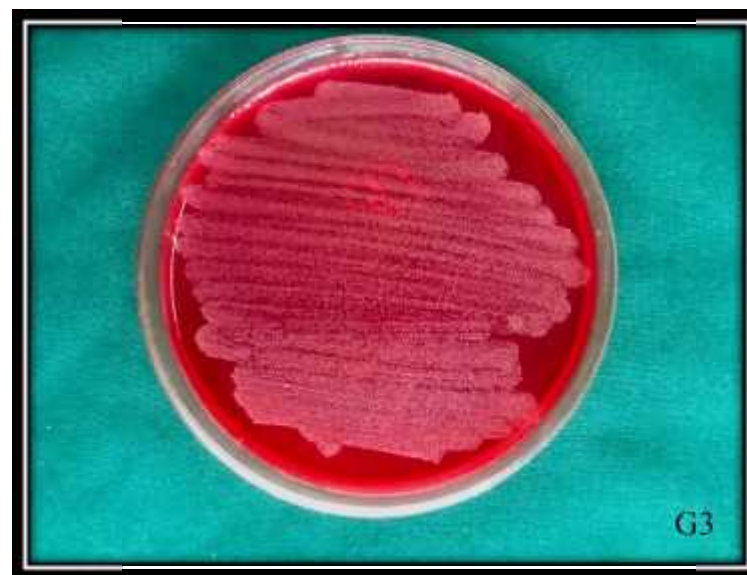
**Fig 18:** Test samples inoculated with *E. faecalis*



**Fig 19:** Pre-treatment *E. faecalis* colonies of G1



**Fig 20:** Pre-treatment *E. faecalis* colonies of G2



**Fig 21:** Pre-treatment *E. faecalis* colonies of G3



**Fig 22:** Incubator



**Fig 23:** Mixing of the Medicament



**Fig 24:** Medicament placement using lentulospiral



**Fig 25:** Test samples were kept in incubator for 7 days



**Fig 26:** Ultrasonic irrigation to remove the medicament



**Fig 27:** Ultrasonic irrigation to remove the medicament



**Fig 28:** Post medicament colonies seen in Group 1.



**Fig 29:** Post medicament colonies seen in Group 2.



**Fig 30:** Post medicament colonies seen in Group 3.

## RESULTS

### Statistical analysis and results

**Table 1: Normality of MIC and MBC scores in the three groups (1, 2, 3) by Kolmogorov Smirnov test**

Parameters	Group 1		Group 2		Group 3	
	Z-value	P-value	Z-value	P-value	Z-value	P-value
MIC	1.2430	0.0910	1.2430	0.0910	0.7980	0.5470
MBC	1.0680	0.2040	1.2430	0.0910	0.7980	0.5470

The MIC and MBC scores in the three groups (1, 2, 3) follow a normal distribution. Therefore, the one way ANOVA and followed by Tukeys multiple posthoc procedures is used.

**Table 2: Summary statistics of MIC scores in the three groups (1, 2, 3)**

Groups	N	Min	Max	Mean	SD	SE	95% CI for mean	
							Lower bound	Upper bound
Group 1	9	25.00	50.00	33.33	12.50	4.17	23.73	42.94
Group 2	9	12.50	25.00	16.67	6.25	2.08	11.86	21.47
Group 3	9	12.50	50.00	29.17	16.54	5.51	16.46	41.88

**Table 3: Comparison of the three groups (1, 2, 3) with mean MIC scores by one way ANOVA**

Sources of variation	Degrees of freedom	Sum of squares	Mean sum of squares	F-value	p-value
Between groups	2	1354.17	677.08	4.3333	0.0247*
Within groups	24	3750.00	156.25		
Total	26	5104.17			

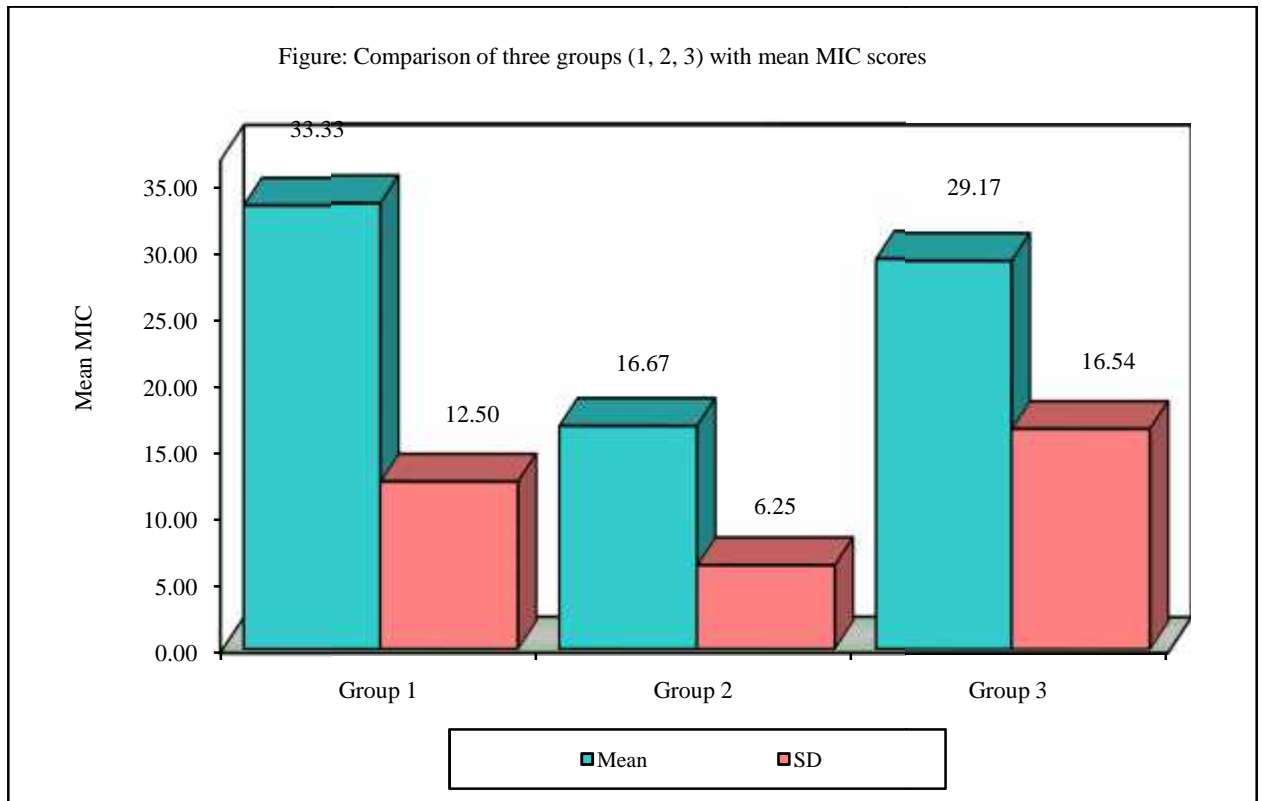
\*p<0.05

**Table 4: Pairwise comparison of the three groups (1, 2, 3) with mean MIC scores by Tukeys multiple posthoc procedure**

Groups	Group 1	Group 2	Group 3
Mean	33.33	16.67	29.17
SD	12.50	6.25	16.54
Group 1	-		
Group 2	P=0.0244*	-	
Group 3	P=0.7618	P=0.1067	-

\*p<0.05

**Figure 1: Comparison of the three groups with mean MIC scores**



**Table 5: Summary of MBC scores in the three groups (1, 2, 3)**

Groups	N	Min	Max	Mean	SD	SE	95% CI for mean	
							Lower bound	Upper bound
Group 1	9	50.00	100.00	72.22	26.35	8.78	51.97	92.48
Group 2	9	25.00	50.00	33.33	12.50	4.17	23.73	42.94
Group 3	9	25.00	100.00	58.33	33.07	11.02	32.91	83.75

**Table 6: Comparison of the three groups (1, 2, 3) with mean MBC scores by one way ANOVA**

Sources of variation	Degrees of freedom	Sum of squares	Mean sum of squares	F-value	p-value
Between groups	2	6990.74	3495.37	5.3929	0.0116*
Within groups	24	15555.56	648.15		
Total	26	22546.30			

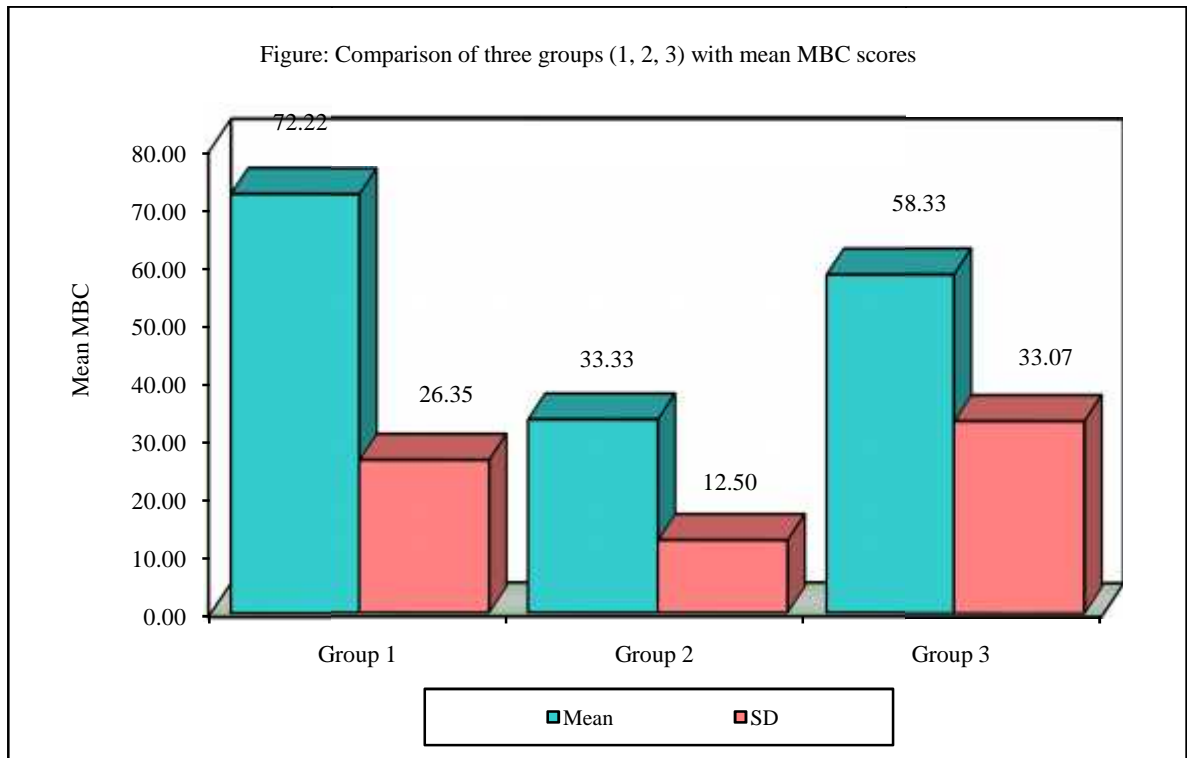
\*p<0.05

**Table 7: Pairwise comparison of the three groups (1, 2, 3) with mean MBC scores by Tukey’s multiple posthoc procedure**

Groups	Group 1	Group 2	Group 3
Mean	72.22	33.33	58.33
SD	26.35	12.50	33.07
Group 1	-		
Group 2	P=0.0095*	-	
Group 3	P=0.4895	P=0.1147	-

\*p<0.05

**Figure 2: Comparison of the three groups with mean MBC scores**



**Table 8: Normality of before and after treatment CFU counts in three groups (1, 2, 3) by Kolmogorov Smirnov test**

	Group 1		Group 2		Group 3	
	Z-value	P-value	Z-value	P-value	Z-value	P-value
Before	0.7660	0.6010	0.7170	0.6830	0.7370	0.6500
After	0.4810	0.9750	-	-	0.4630	0.9830
Difference	0.7660	0.6010	0.7170	0.6830	0.7360	0.6500

The before and after treatment CFU counts in three groups (1, 2, 3) follow a normal distribution. Therefore, the one way ANOVA and followed by Tukey’s multiple posthoc procedure is used.

**Table 9: Summary statistics of before and after treatment CFU counts in the three groups (1, 2, 3)**

Treatment	Groups	N	Min	Max	Mean	SD	SE	95% CI for mean	
								Lower bound	Upper bound
Before	Group 1	20	220000	270000	243000	17502	3914	234809	251191
	Group 2	20	220000	270000	243000	18382	4110	234397	251603
	Group 3	20	220000	270000	251500	16944	3789	243570	259430
After	Group 1	20	10.0	20.0	15.4	3.3	0.7	13.8	16.9
	Group 2	20	0.0	0.0	0.0	0.0	0.0	0.0	0.0
	Group 3	20	50.0	59.0	54.8	2.6	0.6	53.5	56.0
Change	Group 1	20	219984	269983	242985	17501	3913	234794	251175
	Group 2	20	220000	270000	243000	18382	4110	234397	251603
	Group 3	20	219945	269950	251445	16944	3789	243515	259375

**Table 10: Comparison of three groups (1, 2, 3) with mean before and after treatment CFU counts by one way ANOVA**

Treatment	Sources of variation	Degrees of freedom	Sum of squares	Mean sum of squares	F-value	p-value
Before	Between groups	2	963333333	481666667	1.5516	0.2207
	Within groups	57	17695000000	310438596		
	Total	59	18658333333			
After	Between groups	2	31903.63	15951.82	2703.6977	0.0001*
	Within groups	57	336.30	5.90		
	Total	59	32239.93			
Change	Between groups	2	952694904	476347452	1.5345	0.2243
	Within groups	57	17694667336	310432760		
	Total	59	18647362240			

\*p<0.05

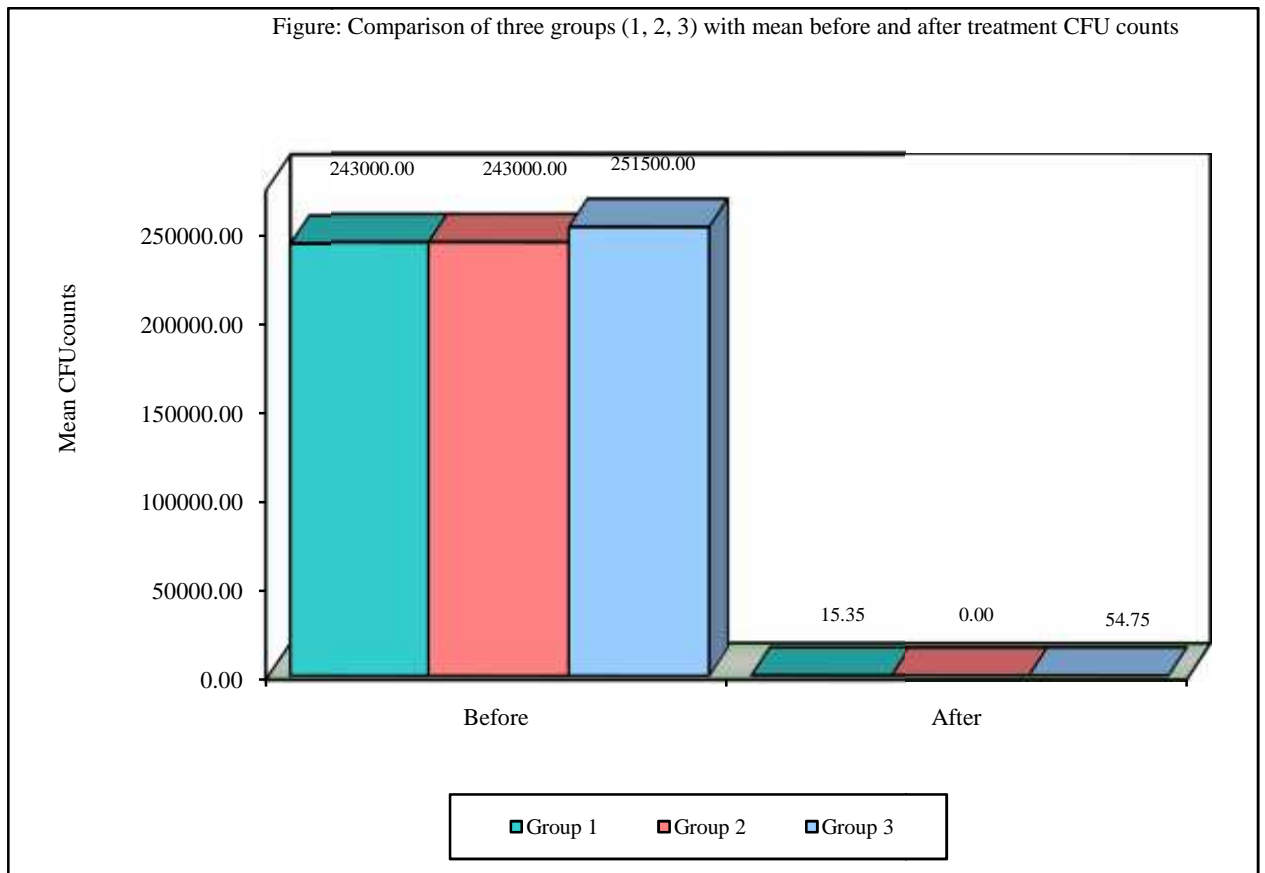
**Table 11: Pairwise comparison of three groups (1, 2, 3) with mean CFU scores by Tukeys multiple posthoc procedure**

Treatment	Groups	Group 1	Group 2	Group 3
Before	Mean	243000	243000	251500
	SD	17502	18382	16944
	Group 1	-		
	Group 2	P=1.0000	-	
	Group 3	P=0.2868	P=0.2868	-
After	Mean	15.35	0.00	54.75
	SD	3.28	0.00	2.63
	Group 1	-		
	Group 2	P=0.0001*	-	
	Group 3	P=0.0001*	P=0.0001*	-
Change	Mean	242985	243000	251445
	SD	17501	18382	16944
	Group 1	-		
	Group 2	P=1.0000	-	
	Group 3	P=0.2900	P=0.2913	-

\*p<0.05

Figure 3: Comparison of three groups with mean before and after treatment

CFU count

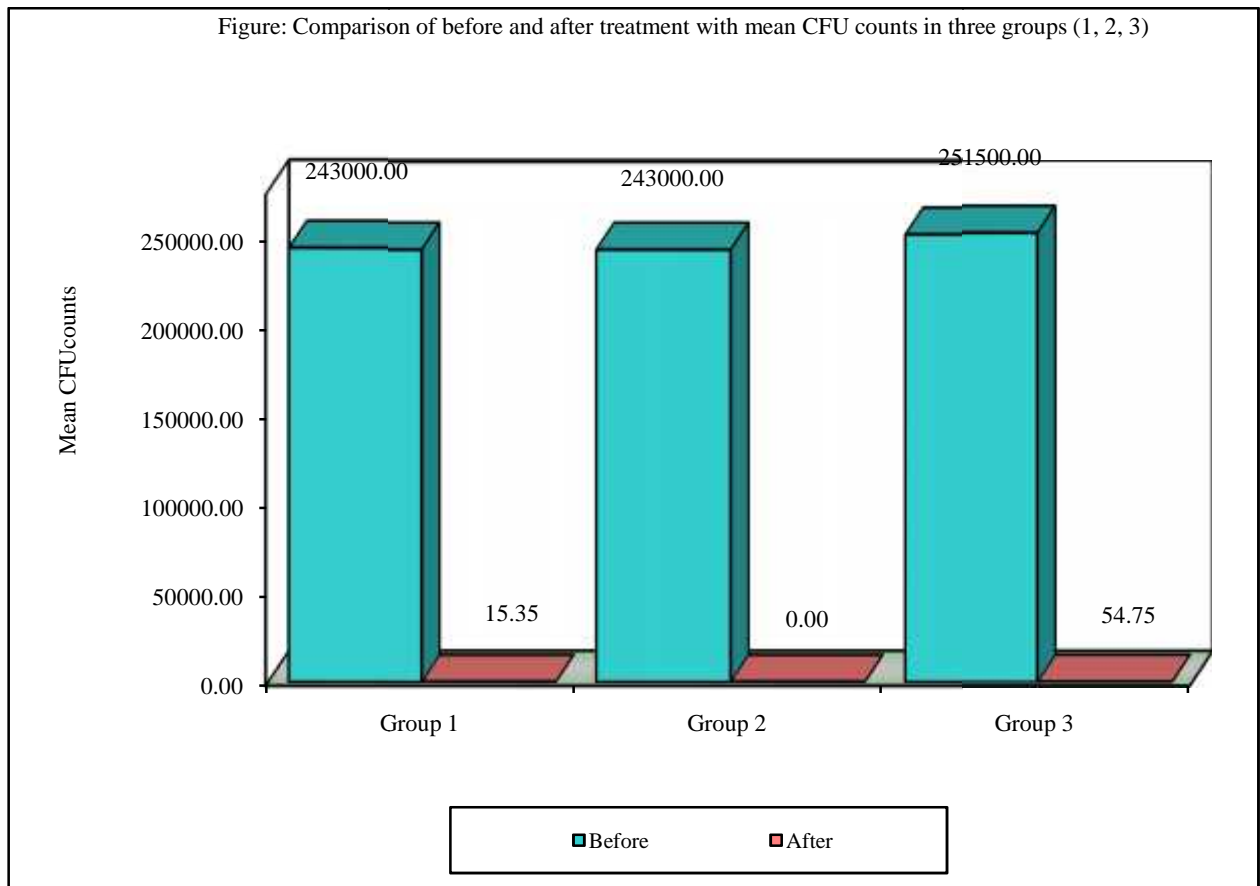


**Table 12: Comparison of before and after treatment with mean CFU counts in three groups (1, 2, 3) by Dependent t-test**

	Treatment	Mean	Std.Dv.	Mean Diff.	SD Diff.	Paired t	p-value
Group 1	Before	243000.00	17501.88				
	After	15.35	3.28	242984.65	17501.13	62.0909	0.0001*
Group 2	Before	243000.00	18381.91				
	After	0.00	0.00	243000.00	18381.91	59.1195	0.0001*
Group 3	Before	251500.00	16944.18				
	After	54.75	2.63	251445.25	16944.44	66.3638	0.0001*

\*p<0.05

**Figure 4: Comparison of before and after treatment with a mean CFU count in the three groups**



**Results:**

The following hypothesis was taken for this study-

**H0:** The null hypothesis taken was that there is no difference in the antibacterial efficacy of Calcium Hydroxide, Diclofenac in combination with Calcium Hydroxide and Ibuprofen used in combination with Calcium Hydroxide when used as an intra-canal medicament.

**H1:** There is a difference in the antibacterial efficacy of Calcium Hydroxide, Diclofenac in combination with Calcium Hydroxide and Ibuprofen in combination with Calcium Hydroxide when used as an intra-canal medicament.

After checking for normal distribution of the tests, further statistical analysis was done to test the significance of the interaction in each of the groups.

The one way ANOVA has done using the MIC and MBC (Table 3 and Table 6) have shown statistically significant results ( $p < 0.05$ ). Since there is less than 5% probability of the success of the null hypothesis, the null hypothesis (H0) is rejected and the alternate hypothesis (H1) is accepted.

Among the three groups, pairwise tests by Tukey's multiple posthoc procedure (Table 4, Table 7 and Table 11) has shown that Group 2 is the most efficient when compared to Group 1 and Group 3 in terms of MIC, MBC and CFU counts ( after treatment efficacy).

One way ANOVA performed using CFU counts for before and after treatment using medicament shows that after medicament is statistically significant with  $P = 0.0001$  (Table 10). Hence H0 is rejected and H1 is accepted i.e there is a difference

in the antibacterial efficacy of the three groups when used as an intra-canal medicament.

Tukeys multiple posthoc procedure is done using CFU mean counts, shows that post-treatment mean CFU count is statistically significant among the three groups 1, 2, 3. In post-treatment CFU counts among the 3 groups, **group 2** has the lowest mean CFU count (0.00) compared to group 1 and group 3 ( 15.35 and 54.75 respectively).

Therefore, in this study, **group 2** i.e. Diclofenac in combination with Calcium Hydroxide is accepted as having better antibacterial effectiveness when used as an intracanal medicament.

## DISCUSSION

Anatomical irregularities are one of the prime factors for the harbouring of microorganisms and necrotic tissue debris, even after meticulous instrumentation and chemical disinfection<sup>46</sup>. Literature shows that in routine cases of an infected root canal, the CFU counts are in the range of  $10^2 - 10^8$ <sup>47</sup>. Progressive enlargement of the canal was suggested by Dalton et al. for the reduction of bacterial colonies, however, the subsequent reduction obtained was only 28% of the total bacterial eradication<sup>48</sup>.

Shuping et al. has reported bacterial reduction up to 61.9% with the usage of NaOCl and rotary instrumentation<sup>49</sup>. Law et al. has reported the re-emergence of bacterial level the same as that of pre-treatment in an instrumented unfilled canal within 2-4 days. In such situations, the usage of intracanal medicament is recommended<sup>50</sup>.

*E. faecalis* an anaerobic facultative microorganism was taken for this study, due to its higher correlation in cases with re-infection. Furthermore, Baumgardener et al. reported in his study that when apical 5mm of the root was exposed to caries it was found that 34% of the bacteria found were facultative anaerobes<sup>51</sup>. Studies have shown the ability of *E. faecalis* to persist during chemo-mechanical preparation and also to resist the high pH of calcium hydroxide due to the presence of proton pump<sup>52</sup>.

Despite calcium hydroxide being the gold standard for intracanal medicament, a recent study done by Tanriverdi et al. and Lana et al., shows less effectiveness i.e only 70% bacterial elimination of *E. faecalis*<sup>53,54</sup>. Several medicaments such as chlorhexidine<sup>55,56,57-58</sup> iodine<sup>59</sup>, camphorated paramonochlorophenol<sup>60, 61</sup>, iodoform<sup>62</sup>,

propolis<sup>63</sup>, and linezolid<sup>64</sup> are added to overcome the drawbacks of calcium hydroxide.

The first antibacterial efficacy of NSAIDs with sodium salicylate was reported by Domenico et al. against *Klebsiella pneumonia*<sup>65</sup>. Recent studies<sup>66-69</sup> through an *In-vitro* and *In-vivo* animal study concluded that Diclofenac has anti-bacterial efficacy against *Salmonella typhimurium*, *Mycobacterium tuberculosis*, and *Listeria monocytogenes*. Moreover, Shirin et al. emphasized the usage of Ibuprofen as an antibacterial medicament against *Helicobacter pylori*<sup>70</sup>. Recently Freitas et al. proved increased anti-bacterial efficacy of 'NSAIDs' with Calcium Hydroxide on a bovine dentin block. Therefore, the potential advantages of combining NSAIDS with Calcium Hydroxide as an intracanal medicament are anti-inflammatory action, local analgesia, and possible antibacterial action with added benefits of Calcium Hydroxide<sup>71</sup>.

The MIC results confining to the study were tested using the tube dilution method, whereas the MBC of the test medicament was detected by sub-culturing these on a medicament free media. The advantage of using this technique is that the examination of a substantial number of bacterial cells could be done by a large initial inoculum provided by the relatively large volume of broth in each of the tubes. The principal disadvantages of the same include the tedious manual task of preparing the two-fold solutions for each test, along with relatively large amounts of reagents and space required for such tests. The test also results in the possibility of making errors in the preparation of medicament concentrations<sup>72</sup>.

The inhibition produced by different brands of a medicament against a particular organism depends upon various extrinsic and intrinsic factors. As per Salem-Milani et al. the effective drug dose (MIC) of Diclofenac and Ibuprofen against *E. faecalis* is 50 µg/ml and above (RW). Similarly, Blanscet et al. showed antibacterial activity of Ca(OH)<sub>2</sub> at 400 µg/ml and 600 µg/ml<sup>99</sup>. In this study, the mean MIC [Tab.2] against *E.faecalis* for Diclofenac with Ca(OH)<sub>2</sub> group was 16.6µg/l, whereas Ibuprofen with Ca(OH)<sub>2</sub> group was 29.17µg/l. The mean MBC [Tab.5] for Diclofenac with Ca(OH)<sub>2</sub> group was 33.3µg/l and Ibuprofen with Ca(OH)<sub>2</sub> group was 58.3µg/l. Also according to the present study, both MIC(33.3 µg/l) and MBC(72.2 µg/l) results of Ca(OH)<sub>2</sub> came out to be less effective than both, group 2(Diclofenac with Ca(OH)<sub>2</sub>) and group 3(Ibuprofen with Ca(OH)<sub>2</sub>).

In the present study, single-rooted, single canal, extracted maxillary central incisors were included to eliminate any anatomical complexities which would lead to any bias in the study<sup>74</sup>.

To eliminate any variable in access preparation, the root lengths were adjusted to 14 mm by sectioning using a high-speed diamond bur and water spray<sup>40</sup>. This was also done to establish a level surface that would serve as a stable and equivocal reference for all measurements.

ProtaperNiTi rotary instruments were used to prepare the canal over stainless steel instruments because NiTiProtaper instruments are designed to cover the whole range of treatment with only a few sets of files. The file system has superior flexibility, unmatched efficiency, and improved safety<sup>75</sup>. The apical canal preparation was done to size F3/F4 depending upon the apical constriction of maxillary central incisors<sup>40</sup>.

In the debridement and disinfection of the root canal system, irrigation plays a primary role. For the present study, 3% sodium hypochlorite (NaOCl), 17% Ethylene diaminetetracetic acid (EDTA), and 5 ml of distilled water were used according to the following protocol<sup>76</sup>.

- In between each instrumentation: 2ml of 3% NaOCl per canal
- After shaping: 5 ml of 3% NaOCl followed by 5 ml of 17% EDTA per canal.
- After preparation, the canals were irrigated with 5 ml of distilled water and dried with sterile paper points.

For the present study, 3% sodium hypochlorite (Vishal DentocarePvt Ltd., India) was used because of its effective antimicrobial and tissue dissolution property<sup>77</sup>. Studies have shown, for removal of vital and necrotic tissue concentrations of NaOCl above 2% have the dissolving characteristics. Even at lower concentrations NaOCl has strong antimicrobial activity (Bystrom and Sundquist 1983) and kills bacteria very rapidly<sup>78</sup>.

17% EDTA solution (Canalarge, Ammdent, Chandigarh, India) was used as an irrigant because it removes the debris by chelating the inorganic tissue. The smear layer refers to this debris, which includes soft tissue and dentine components and the former interrupts the penetration of the irrigation (Violich and Chandler 2010). NaOCl is used along with EDTA to increase the antibacterial effectiveness when used in the same irrigating regime. (Bystrom and Sundqvist 1985)<sup>79</sup>.

5ml of distilled water (NICE LIFECARE, NEW DELHI) was used as a final flush to neutralize various chemicals of the irrigants by its flushing action<sup>80</sup>.

To make the negotiation and movement of the file within the root easier 17% EDTA gel (Well Prep, Vericom, Korea), was used in this study as a lubricant to facilitate instrumentation of the root canal. It was used with stainless steel and nickel-titanium file systems in the early stages of canal negotiation and shaping<sup>81</sup>.

Needle irrigation is the most common method for the delivery of irrigants into the root canal system. Large syringes can be used to deliver a large volume of irrigant, but these are difficult to control<sup>82</sup>. A study done by Gopikrishna et al. concluded that a 26-gauge needle provides a higher flow rate, but impairs the depth of penetration and a 30-gauge needle had the slowest flow rate. Hence a 5ml side vented syringe with a 27 gauge needle, which has a good depth of penetration as well as flow rate was used in study<sup>82</sup>.

The endodontic model was inoculated with the test organism *E.faecalis* for 2 weeks. This *In-vitro* model stimulates an actual clinical situation resembling endodontic retreatment. The inoculation time for the adequate growth of the bacteria was kept under 2 weeks because it is difficult to maintain the viability of the test organism in an *In-vitro study*<sup>83</sup>. The usage of an endodontic model gives us a better understanding of dentin buffering action, which otherwise would not have been possible, had the study been done on a Petri dish<sup>84</sup>.

A 4 mm thick coronal seal was placed to restrict the nutrient supply to the test organism<sup>85</sup>. The medicaments were applied for seven days to check their antimicrobial effect. It was observed by Sjogren et al. that a negative culture was attainable with the placement of a Ca(OH)<sub>2</sub> for just 7 days<sup>86</sup>.

In this study, a leptulospiral was used for the placement of intracanal medicament. The insertion of the paste was performed using small quantities of the medicament at a time. Lentulospiral propels the medicament centrifugally and helps in better placement<sup>87</sup>.

Passive ultrasonic agitation of irrigation solutions is more effective than syringe irrigant alone. The hydrodynamic phenomenon caused by vibration of the file in the canal filled with irrigant creates acoustic microstreaming. This microstreaming is responsible for the removal of root canal debris<sup>88</sup>. Therefore, PUI was also used for 3 minutes in the study for the removal of intracanal medicaments.

For the beneficial effect of the intracanal medicament, it is of utmost importance that the medicament to come in direct contact with the microorganism. Studies have shown that the penetration of the medicament is directly proportional to the penetration and solubility of the drug<sup>89</sup>. The complexation method was used to make the drugs water-soluble to increase their penetration in the dentinal tubules. The medicaments were prepared by mixing them with cyclodextrin in a 1:1 ratio. cyclodextrin ( -1,4)-linked oligosaccharides of -d-glucopyranose is a solubility enhancer which clathrates the test medicament and helps in better dissolution in distilled water<sup>84</sup>.

Instrumentation and microbiological analyses were done by a single operator, to eliminate inter-operator bias. The samples were attained by the circumferential pumping action of the file, to get a suspension of bacteria from root canal walls, dentin, and the apical delta. The pumping action also aids in the disruption of biofilms in these areas<sup>74</sup>.

In the present study, thioglycolate broth was used to keep the viability of the microorganism during transportation. The sampling was repeated using additional paper points until all the glucose broth was absorbed. According to Fouad et al. the last paper point is most important because it will absorb the liquid from the most peripheral areas of the apical region <sup>74</sup>.

In the present study, the first sample (S1) reading of all the three groups was in the range of  $2.2 \times 10^6$  -  $2.7 \times 10^6$ , marking the same CFU count in all the three groups. Post medicament it was observed that all the groups (Group I, Group II, Group III) showed reduced CFU counts, therefore the antibacterial effect was significant in all of them [ Fig 3].

Comparing the medicaments with the control group  $\text{Ca(OH)}_2$  (Group-1), Diclofenac with  $\text{Ca(OH)}_2$  (Group-2) and Ibuprofen with  $\text{Ca(OH)}_2$  (Group-3) showed very strong evidence against the null hypothesis ( $P = 0.0001$ ) [Figure 3][Tab.10].

This is in favour of earlier research done by de Freitas et al. ( 2017) which documented enhanced antibacterial activity of  $\text{Ca(OH)}_2$  in combination with NSAIDs <sup>39</sup>. But the present findings were not in accordance with their study in terms of the increased antibacterial efficacy of Ibuprofen in combination with  $\text{Ca(OH)}_2$ . This may be because Ibuprofen is lipophilic <sup>101</sup> i.e. it is not water-soluble as opposed to diclofenac which is water-soluble, [albeit sparingly] <sup>101</sup>.

Besides when Diclofenac with Calcium Hydroxide (Group 2) was compared with Ibuprofen with  $\text{Ca(OH)}_2$  group 3 also showed strong evidence against the null hypothesis [ $p < 0.05$ ]. Thus, the null hypothesis was rejected.

Salem-Milani et al. , considered the first one to ever study the antibacterial efficiency of Ibuprofen, Diclofenac, and Ca(OH)<sub>2</sub> by utilizing the agar diffusion test against *E. faecalis* and concluded that NSAIDs can be used as an intracanal medicament<sup>37</sup>.

The results of this study partly concur with a study by Salem-Milani et al., where a greater antibacterial action was shown by diclofenac and Ibuprofen than Ca(OH)<sub>2</sub>.

In the present study when the intergroup i.e group 1(Ca(OH)<sub>2</sub>), group 2(Diclofenac +Ca(OH)<sub>2</sub>), group 3(Ibuprofen+ Ca(OH)<sub>2</sub>) were compared strong evidence against the null hypothesis [p<0.05] was proven. Therefore once again, the null hypothesis was rejected. Several studies have documented the antibacterial activity of Diclofenac and Ibuprofen but the exact mechanism remains uncertain. However, studies have proposed the following mechanism(s) of action:

- Inhibition of bacterial DNA synthesis<sup>69</sup>
- Impairment of membrane activity<sup>67,68</sup>
- Anti-plasmid activity<sup>101</sup>
- Alteration in genes encoding transport/binding proteins,
- DNA synthesis and cell envelope<sup>100</sup>
- Down-regulation of efflux pumps<sup>100</sup>
- Reduced quorum sensing-controlled motility leading to reduced biofilm<sup>102</sup>.

Siqueira and Uzeda et al. demonstrated that 1 week of contact with Ca(OH)<sub>2</sub> mixed with saline had limited effect in *E. faecalis* eradication<sup>91</sup>. Estrela et al. reported in her study that Ca(OH)<sub>2</sub> had no antimicrobial effect on *E. faecalis*, *S. aureus*,

*Bacillus subtilis*, *Pseudomonas aeruginosa*<sup>92</sup>. In a Systematic review to assess the antibacterial efficacy of Ca(OH)<sub>2</sub> by using culture techniques, Sathorn et al. reported that Ca(OH)<sub>2</sub> had decreased efficacy in eradicating bacteria<sup>93</sup>.

The probable mechanism for the antimicrobial action of Calcium Hydroxide is the release of hydroxyl ions in an aqueous environment which causes damage to the bacterial cytoplasmic membrane and also protein denaturation leading to the damage of DNA<sup>90</sup>.

The limited antimicrobial effect of Calcium Hydroxide on facultative anaerobes could be attributed to the dentine buffering effect. The antibacterial activity of Calcium Hydroxide is driven by its high pH (12.5) due to which it has a destructive impact on bacterial cell membranes and protein structure. For medicament's efficacy, the hydroxyl ions from Calcium Hydroxide must penetrate and reach dentin sufficiently. This diffusion is made difficult since dentin hydroxyapatite has a strong buffering property that must be overcome by the hydroxyl ions leaving the root canal space. The buffering of alkaline substances in the dentin is due to the proton donors in the hydroxyl layer of the hydroxyapatite. It is seen that Calcium Hydroxide alkalizes the dentin, even so, it is unable to eradicate *E.faecalis*, which survives high pH up to 11.5<sup>94</sup>.

One of the few mechanisms that explain the poor action of Calcium Hydroxide on *E.faecalis* is the ability of *E.faecalis* to penetrate deep into the dentin to the depth of 1000µm<sup>95</sup> which facilitates its protection from chemomechanical root canal preparations. Another possible mechanism is the formation of co-aggregates with other bacterial species.

Also, *E.faecalis* biofilms have exhibited the ability to calcify, which may further facilitate their stability. Studies have also shown that the virulence factors within the *E.faecalis* have the potential to adapt and survive, due to the presence of the transcripts of *ftsZ*, a gene involved in cell division that helps its multiplication.<sup>96,97</sup> The presence of the above factors along with many other virulence traits such as enterococcus surface protein, a collagen-binding protein with the secretion of proteases (ex. Gelatinase) and lexin (ex. cytolysin) gives it an upper hand over the other bacterial species found in the root canal, all of which contributes to its survival.<sup>98</sup>

In this study, medicaments in their powder forms were mixed in 1:1 ratio (approx.) with distilled water. This was because of the difficulty in measuring such a small amount of the medicament as well as due to the complexity in delivering the medicament inside the root canal<sup>38</sup>.

The complexation procedure which was done to enhance the solubility of the medicament did indeed enhance the solubility to an extent but didn't make the drug completely soluble<sup>40</sup>. It is well documented that the penetration is directly dependent on the solubility, which in turn helps the medicament to come in direct contact with the bacteria. Therefore in the present study, Ibuprofen with Calcium Hydroxide did not show comparable results as Diclofenac with Calcium Hydroxide which showed excellent results.

According to Sjögren et al, a 7-day placement of a  $\text{Ca(OH)}_2$  was adequate in complete eradication of the root canal bacteria. Similarly, Shuping et al in 2000 reported that 92.5% of root canals showed negative culture in a 1 week after

placement of Ca(OH)<sub>2</sub>. Prolong duration of placement of intra-canal medicament (Ca(OH)<sub>2</sub>) causes has a negative effect on fracture resistance of the teeth.

The present study shows that the combination of diclofenac with Ca(OH)<sub>2</sub> is effective in complete eradication of root canal bacteria in 7 days, and thus minimizes any damage to the root fracture resistance of the treated tooth.

Therefore Diclofenac with Ca(OH)<sub>2</sub> with both anti-inflammatory and antibacterial activity can be a potential intracanal medicament and can be a replacement for Calcium Hydroxide with further research.

Further *In vivo* studies are to be carried out for the practical implementation in the clinical scenario of results obtained from the present study.

The incompetent testament of Ibuprofen could be attributed to the medicament preparation at a molecular level that might be one of the reasons for the lack of synergistic effect with Ca(OH)<sub>2</sub>. Also, appropriate research should be carried out on the proportionating field of the combination of the medicament so that we can get the practically achievable value of the medicament that can be used in a clinical scenario. In our further research, we would want to study if this combination could decrease the placement time of intracanal medicament so that fracture resistance of the teeth can be maintained.

## CONCLUSION

Under the limitations of the present research, it can be concluded that:

- 1) A combination of NSAIDS with Calcium Hydroxide is effective against *E.faecalis*.
- 2) Diclofenac with Calcium Hydroxide shows 100 % efficacy against *E.faecalis* and can be used in a clinical scenario.
- 3) Ibuprofen with Calcium Hydroxide could not eliminate *E.faecalis* due to decreased solubility and possibly less penetration into the dentinal tubules, therefore further research should be carried to improve the drug at the molecular level.
- 4) The better efficacy of all the combination could be attributed to the complexation method.

## SUMMARY

The study was conducted in the Department of Conservative Dentistry and Endodontics, ViswanathKatti Institute of Dental Sciences, KAHER Belagavi with the aim for comparisons of anti-bacterial efficacy of Calcium Hydroxide, Diclofenac with Calcium Hydroxide and Ibuprofen with Calcium Hydroxide against *Enterococcus faecalis* in an endodontic model.

Bacteria play a pivotal role in the pathogenesis of apical periodontitis. For optimal treatment outcome complete elimination of bacteria is of utmost importance. Cleaning and shaping along with antimicrobial agents lead to a reduction in the bacterial colonies present in the root canal. However, studies have shown chemo-mechanical instrumentation alone is unable to abolish the full bacterial colonies in the root canal. To overcome this, intracanal medication is advocated. Despite favorable clinical evidence of commonly used  $\text{Ca(OH)}_2$ , it has been reported that there is a correlation between  $\text{Ca(OH)}_2$  placement and increased incidence of tooth fracture, principally reported in teeth with an open apex. This led to the addition of newer medicament which could enhance the property of  $\text{Ca(OH)}_2$  and act synergistically to them.

The conventionally used class of analgesics- Non-steroidal anti-inflammatory drugs (NSAIDs)- may possess additional therapeutic properties such as anti-bacterial efficacy through inhibition of bacterial DNA synthesis or impairment of membrane activity; prevention of bacterial colonization and biofilm formation by interfering with quorum sensing of bacteria; and anti-plasmid activity.

Thus, the present investigation was undertaken to evaluate and compare the antibacterial efficacy of  $\text{Ca(OH)}_2$ , Diclofenac with  $\text{Ca(OH)}_2$  and Ibuprofen with  $\text{Ca(OH)}_2$  against *Enterococcus faecalis*.

Sixty extracted human maxillary central incisors teeth were chosen which fulfilled the inclusion and exclusion criteria. De-coronation of samples was done at CEJ using a diamond disc with continuous water coolant for standardization. Biomechanical preparation was done using Protaper Universal NiTi files up to F3/F4. 2ml of 3% of NaOCl was irrigated between each instrument. After completion of chemo-mechanical preparation, sample S-1 was collected by introducing a sterile paper point. The paper point was immediately placed in test tubes containing glucose broth. Intracanal medicaments were placed according to the respective group Group1- $\text{Ca(OH)}_2$ , Group 2- Diclofenac +  $\text{Ca(OH)}_2$ , Group 3- Ibuprofen +  $\text{Ca(OH)}_2$ . Intracanal medicaments were removed after 7 days by using passive ultrasonic irrigation and sample S-2 was collected.

The results were analyzed statistically by One way ANOVA, Tukey's multiple post hoc, Dependent t-test all of which indicated that there is a significant difference between all the three groups (p-value <0.05). Among all the three groups, Group 2 (Diclofenac +  $\text{Ca(OH)}_2$ ) performed better, this could be because of the better synergistic action of both the medicament.

Therefore, the null hypothesis that there is **no difference** in the antibacterial efficacy of Calcium Hydroxide, Diclofenac in combination with Calcium Hydroxide and Ibuprofen in combination with Calcium Hydroxide was rejected.

**BIBLIOGRAPHY**

1. Ørstavik D. Root canal disinfection: a review of concepts and recent developments. *Australian Endodontic Journal*. 2003 Aug;29(2):70-4.
2. Card SJ, Sigurdsson A, Ørstavik D, Trope M. The effectiveness of increased apical enlargement in reducing intracanal bacteria. *Journal of Endodontics*. 2002 Nov 1;28(11):779-83.
3. McGurkin-Smith R, Trope M, Caplan D, Sigurdsson A. Reduction of intracanal bacteria using GT rotary instrumentation, 5.25% NaOCl, EDTA, and Ca (OH) 2. *Journal of Endodontics*. 2005 May 1;31(5):359-63.
4. Zandi H, Rodrigues RC, Kristoffersen AK, Enersen M, Mdala I, Ørstavik D, Rôças IN, SiqueiraJr JF. Antibacterial effectiveness of 2 root canal irrigants in root-filled teeth with infection: a randomized clinical trial. *Journal of Endodontics*. 2016 Sep 1;42(9):1307-13.
5. Rodrigues RC, Zandi H, Kristoffersen AK, Enersen M, Mdala I, Ørstavik D, Rôças IN, SiqueiraJr JF. Influence of the apical preparation size and the irrigant type on bacterial reduction in root canal–treated teeth with apical periodontitis. *Journal of Endodontics*. 2017 Jul 1;43(7):1058-63.
6. Gazzaneo I, Vieira GC, Pérez AR, Alves FR, Gonçalves LS, Mdala I, SiqueiraJr JF, Rôças IN. Root canal disinfection by single-and multiple-instrument systems: Effects of sodium hypochlorite volume, concentration, and retention time. *Journal of endodontics*. 2019 Jun 1;45(6):736-41.
7. Shuping GB, Ørstavik D, Sigurdsson A, Trope M. Reduction of intracanal bacteria using nickel-titanium rotary instrumentation and various medications. *Journal of endodontics*. 2000 Dec 1;26(12):751-5.

8. Shuping GB, Ørstavik D, Sigurdsson A, Trope M. Reduction of intracanal bacteria using nickel-titanium rotary instrumentation and various medications. *Journal of endodontics*. 2000 Dec 1;26(12):751-5.
9. Desai S, Chandler N. Calcium hydroxide-based root canal sealers: a review. *Journal of endodontics*. 2009 Apr 1;35(4):475-80.
10. Özdemir MB, Karata E, Albayrak M, Bayır Y. Effect of intracanal medicaments on matrix metalloproteinase-9 and vasoactive intestinal peptide secretion in periapical lesions of re-treated canals: a randomized controlled clinical study. *Clinical Oral Investigations*. 2019 Feb 8;23(2):921-8.
11. Baumgartner C, Siqueira J, Sedgley CM, Kishen A. Microbiology of endodontic disease. *Endodontics*. Ingle JI, Bakland LK, Baumgartner JC editors. 2008;6:258.
12. Sjögren U, Figdor D, Persson S, Sundqvist G. Influence of infection at the time of root filling on the outcome of endodontic treatment of teeth with apical periodontitis. *International endodontic journal*. 1997 Sep;30(5):297-306.
13. Waltimo T, Trope M, Haapasalo M, Ørstavik D. Clinical efficacy of treatment procedures in endodontic infection control and one year follow-up of periapical healing. *Journal of endodontics*. 2005 Dec 1;31(12):863-6.
14. Louwakul P, Saelo A, Khemaleelakul S. Efficacy of calcium oxide and calcium hydroxide nanoparticles on the elimination of *Enterococcus faecalis* in human root dentin. *Clinical oral investigations*. 2017 Apr 1;21(3):865-71.
15. Louwakul P, Saelo A, Khemaleelakul S. Efficacy of calcium oxide and calcium hydroxide nanoparticles on the elimination of *Enterococcus faecalis* in human root dentin. *Clinical oral investigations*. 2017 Apr 1;21(3):865-71.

16. Dastidar SG, Ganguly K, Chaudhuri K, Chakrabarty AN. The anti-bacterial action of diclofenac shown by inhibition of DNA synthesis. *International journal of antimicrobial agents*. 2000 Apr 1;14(3):249-51.
17. Annadurai S, Basu S, Ray S, Dastidar SG, Chakrabarty AN. Antibacterial activity of the antiinflammatory agent diclofenac sodium. *Indian journal of experimental biology*. 1998 Jan;36(1):86-90.
18. Dutta NK, Annadurai S, Mazumdar K, Dastidar SG, Kristiansen JE, Molnar J, Martins M, Amaral L. Potential management of resistant microbial infections with a novel non-antibiotic: the anti-inflammatory drug diclofenac sodium. *International journal of antimicrobial agents*. 2007 Sep 1;30(3):242-9.
19. Dutta NK, Mazumdar K, Dastidar SG, Park JH. Activity of diclofenac used alone and in combination with streptomycin against *Mycobacterium tuberculosis* in mice. *International journal of antimicrobial agents*. 2007 Oct 1;30(4):336-40.
20. Ulusoy S, Bosgelmez-Tinaz G. Nonsteroidal anti-inflammatory drugs reduce the production of quorum sensing regulated virulence factors and swarming motility in human pathogen *Pseudomonas aeruginosa*. *Drug research*. 2013 Aug;63(08):409-13.
21. Mazumdar K, Dastidar SG, Park JH, Dutta NK. The anti-inflammatory non-antibiotic helper compound diclofenac: an antibacterial drug target. *European journal of clinical microbiology & infectious diseases*. 2009 Aug 1;28(8):881.
22. Kristiansen JE, Amaral L. The potential management of resistant infections with non-antibiotics. *The Journal of antimicrobial chemotherapy*. 1997 Sep 1;40(3):319-27.

23. Ehrmann EH, Messer HH, Adams GG. The relationship of intracanal medicaments to postoperative pain in endodontics. *International endodontic journal*. 2003 Dec;36(12):868-75.
24. Haapasalo M, Qian W. Irrigants and intracanal medication. *Ingle's Endodontics*. 2008;6:997-1008.
25. Yassen GH, Eckert GJ, Platt JA. Effect of intracanal medicaments used in endodontic regeneration procedures on microhardness and chemical structure of dentin. *Restorative dentistry & endodontics*. 2015 May 1;40(2):104-12.
26. Zehnder M. Root canal irrigants. *Journal of endodontics*. 2006 May 1;32(5):389-98
27. Kim JH, Kim Y, Shin SJ, Park JW, Jung IY. Tooth discoloration of immature permanent incisor associated with triple antibiotic therapy: a case report. *Journal of endodontics*. 2010 Jun 1;36(6):1086-91.
28. Petrino JA, Boda KK, Shambarger S, Bowles WR, McClanahan SB. Challenges in regenerative endodontics: a case series. *Journal of endodontics*. 2010 Mar 1;36(3):536-41.
29. Prather BT, Ehrlich Y, Spolnik K, Platt JA, Yassen GH. Effects of two combinations of triple antibiotic paste used in endodontic regeneration on root microhardness and chemical structure of radicular dentine. *Journal of oral science*. 2014;56(4):245-51.
30. Sponchiado EC, Pereira JV, Marques AA, Garcia LD, França SC. In vitro assessment of antimicrobial activity of Pothomorpheumbellata extracts against *Enterococcus faecalis*. *Indian Journal of Dental Research*. 2014 Jan 1;25(1):64.Orstavik D, editor. *Essential endodontology: prevention and treatment of apical periodontitis*. John Wiley & Sons; 2020 Jan 21.

31. Nigam P, Nath G, Gulati AK. Antibacterial effect of calcium hydroxide in endodontic treatment. *Endodontology*, 1991; 3 (1): 36-40.
32. Lana PE, Scelza MF, Silva LE, Mattos-Guaraldi AL, Hirata Júnior R. Antimicrobial activity of calcium hydroxide pastes on *Enterococcus faecalis* cultivated in root canal systems. *Brazilian dental journal*. 2009;20(1):32-6.
33. Sjögren U, Figdor D, Spångberg L, Sundqvist G. The antimicrobial effect of calcium hydroxide as a short-term intracanal dressing. *International endodontic journal*. 1991 May;24(3):119-25.
34. Kim D, Kim E. Antimicrobial effect of calcium hydroxide as an intracanal medicament in root canal treatment: a literature review-Part I. In vitro studies. *Restorative dentistry & endodontics*. 2014 Nov 1;39(4):241-52.
35. Sathorn C, Parashos P, Messer H. Antibacterial efficacy of calcium hydroxide intracanal dressing: a systematic review and meta-analysis. *International endodontic journal*. 2007 Jan;40(1) Whitbeck ER, Quinn GD, Quinn JB. Effect of calcium hydroxide on the fracture resistance of dentin. *Journal of research of the National Institute of Standards and Technology*. 2011 Jul;116(4):743.:2-10.
36. Whitbeck ER, Quinn GD, Quinn JB. Effect of calcium hydroxide on the fracture resistance of dentin. *Journal of research of the National Institute of Standards and Technology*. 2011 Jul;116(4):743.
37. Salem-Milani A, Balaei-Gajan E, Rahimi S, Moosavi Z, Abdollahi A, Zakeri-Milani P, Bolourian M. Antibacterial effect of diclofenac sodium on *Enterococcus faecalis*. *Journal of dentistry (Tehran, Iran)*. 2013;10(1):16.
38. Chockattu SJ, Deepak BS, Goud KM. Comparison of anti-bacterial efficiency of ibuprofen, diclofenac, and calcium hydroxide against *Enterococcus faecalis* in

- an endodontic model: an in vitro study. *Journal of conservative dentistry: JCD*. 2018 Jan;21(1):80.
39. deFreitas RP, Greatti VR, Alcalde MP, Cavenago BC, Vivan RR, Duarte MA, Weckwerth AC, Weckwerth PH. Effect of the association of nonsteroidal anti-inflammatory and antibiotic drugs on antibiofilm activity and pH of calcium hydroxide pastes. *Journal of endodontics*. 2017 Jan 1;43(1):131-4.
40. Khadka P, Ro J, Kim H, Kim I, Kim JT, Kim H, Cho JM, Yun G, Lee J. Pharmaceutical particle technologies: An approach to improve drug solubility, dissolution and bioavailability. *Asian journal of pharmaceutical sciences*. 2014 Dec 1;9(6):304-16.
41. Karata E, Baltacı MÖ, Uluköylü E, Adıgüzel A. Antibacterial effectiveness of calcium hydroxide alone or in combination with Ibuprofen and Ciprofloxacin in teeth with asymptomatic apical periodontitis: a randomized controlled clinical study. *International Endodontic Journal*. 2020 Jun;53(6):742-53.
42. Alghamdi F, Shakir M. The Influence of *Enterococcus faecalis* as a Dental Root Canal Pathogen on Endodontic Treatment: A Systematic Review. *Cureus*. 2020 Mar;12(3).
43. Sungur DD, Aksel H, Purali N. Effect of a low surface tension vehicle on the dentinal tubule penetration of calcium hydroxide and triple antibiotic paste. *Journal of endodontics*. 2017 Mar 1;43(3):452-5.
44. Holland R, Scares IJ, Scares IM. Influence of irrigation and intracanal dressing on the healing process of dogs' teeth with apical periodontitis. *Dental Traumatology*. 1992 Dec;8(6):223-9. Holland R, Scares IJ, Scares IM. Influence of irrigation and intracanal dressing on the healing process of dogs' teeth with apical periodontitis. *Dental Traumatology*. 1992 Dec;8(6):223-9.

45. Haapasalo M, Udnæs T, Endal U. Persistent, recurrent, and acquired infection of the root canal system post-treatment. *Endodontic topics*. 2003 Nov;6(1):29-56.
46. Peters OA, Schönenberger K, Laib A. Effects of four Ni–Ti preparation techniques on root canal geometry assessed by micro computed tomography. *International endodontic journal*. 2001 Apr;34(3):221-30.
47. Shuping GB, Ørstavik D, Sigurdsson A, Trope M. Reduction of intracanal bacteria using nickel-titanium rotary instrumentation and various medications. *Journal of endodontics*. 2000 Dec 1;26(12):751-5.
48. Byström A, Happonen RP, Sjögren U, Sundqvist G. Healing of periapical lesions of pulpless teeth after endodontic treatment with controlled asepsis. *Dental traumatology*. 1987 Apr;3(2):58-63.
49. Sen A, Batra A. Determination of antimicrobial potentialities of different solvent extracts of the medicinal plant: *Phyllanthusamarus* Schum. and Thonn. *International Journal of Green Pharmacy (IJGP)*. 2012;6(1).
50. Baumgartner JC, Siqueira JF, Sedgley CM, Kishen A. Microbiology of endodontic disease. *Ingle's endodontics*. 2008;6:221-308.
51. Sundqvist G. Taxonomy, ecology, and pathogenicity of the root canal flora. *Oral Surgery, Oral Medicine, Oral Pathology*. 1994 Oct 1;78(4):522-30.
52. Baumgartner JC, Falkler WA. Bacteria in the apical 5 mm of infected root canals. *Journal of endodontics*. 1991 Aug 1;17(8):380-3.
53. Evans M, Davies JK, Sundqvist G, Figdor D. Mechanisms involved in the resistance of *Enterococcus faecalis* to calcium hydroxide. *International endodontic journal*. 2002 Mar;35(3):221-8.

54. Tanriverdi F, Esener T, Erganis O, Belli S. An in vitro test model for investigation of disinfection of dentinal tubules infected with *Enterococcus faecalis*. *Braz Dent J*. 1997 Feb;8(2):67-72.
55. Lana PE, Scelza MF, Silva LE, Mattos-Guaraldi AL, Hirata Júnior R. Antimicrobial activity of calcium hydroxide pastes on *Enterococcus faecalis* cultivated in root canal systems. *Brazilian dental journal*. 2009;20(1):32-6.
56. Schäfer E, Bössmann K. Antimicrobial efficacy of chlorhexidine and two calcium hydroxide formulations against *Enterococcus faecalis*. *Journal of Endodontics*. 2005 Jan 1;31(1):53-6.
57. Ercan E, Dalli M, Dülgergil ÇT. In vitro assessment of the effectiveness of chlorhexidine gel and calcium hydroxide paste with chlorhexidine against *Enterococcus faecalis* and *Candida albicans*. *Oral Surgery, Oral Medicine, Oral Pathology, Oral Radiology, and Endodontology*. 2006 Aug 1;102(2):e27-31.
58. Sirén EK, Haapasalo MP, Waltimo TM, Ørstavik D. In vitro antibacterial effect of calcium hydroxide combined with chlorhexidine or iodine potassium iodide on *Enterococcus faecalis*. *European journal of oral sciences*. 2004 Aug;112(4):326-31.
59. Souza-Filho FJ, Soares AD, Vianna ME, Zaia AA, Ferraz CC, Gomes BP. Antimicrobial effect and pH of chlorhexidine gel and calcium hydroxide alone and associated with other materials. *Brazilian dental journal*. 2008;19(1):28-33.
60. deMagalhãesSilveira CF, Cunha RS, Fontana CE, de Martin AS, de Almeida Gomes BP, Motta RH, da SilveiraBueno CE. Assessment of the antibacterial activity of calcium hydroxide combined with chlorhexidine paste and other intracanal medications against bacterial pathogens. *European Journal of Dentistry*. 2011 Jan;5(1):1.

61. Vianna ME, Gomes BP, Sena NT, Zaia AA, Ferraz CC, Souza Filho FJ. In vitro evaluation of the susceptibility of endodontic pathogens to calcium hydroxide combined with different vehicles. *Brazilian dental journal*. 2005 Dec;16(3):175-80.
62. Rezende GP, Costa LR, Pimenta FC, Baroni DA. In vitro antimicrobial activity of endodontic pastes with propolis extracts and calcium hydroxide: a preliminary study. *Brazilian dental journal*. 2008;19(4):301-5.
63. Pavaskar R, De Ataíde ID, Chalakkal P, Pinto MJ, Fernandes KS, Keny RV, Kamath A. An in vitro study comparing the intracanal effectiveness of calcium hydroxide—and linezolid-based medicaments against *Enterococcus faecalis*. *Journal of endodontics*. 2012 Jan 1;38(1):95-100.
64. Domenico PH, Schwartz SU, Cunha BA. Reduction of capsular polysaccharide production in *Klebsiella pneumoniae* by sodium salicylate. *Infection and Immunity*. 1989 Dec 1;57(12):3778-82.
65. Dastidar SG, Ganguly K, Chaudhuri K, Chakrabarty AN. The anti-bacterial action of diclofenac shown by inhibition of DNA synthesis. *International journal of antimicrobial agents*. 2000 Apr 1;14(3):249-51.
66. Annadurai S, Basu S, Ray S, Dastidar SG, Chakrabarty AN. Antibacterial activity of the antiinflammatory agent diclofenac sodium. *Indian journal of experimental biology*. 1998 Jan;36(1):86-90.
67. Dutta NK, Annadurai S, Mazumdar K, Dastidar SG, Kristiansen JE, Molnar J, Martins M, Amaral L. Potential management of resistant microbial infections with a novel non-antibiotic: the anti-inflammatory drug diclofenac sodium. *International journal of antimicrobial agents*. 2007 Sep 1;30(3):242-9.

68. Dutta NK, Mazumdar K, Dastidar SG, Park JH. Activity of diclofenac used alone and in combination with streptomycin against *Mycobacterium tuberculosis* in mice. *International journal of antimicrobial agents*. 2007 Oct 1;30(4):336-40.
69. Shirin H, Moss SF, Kancherla S, Kancherla K, Holt PR, Weinstein IB, Sordillo EM. Non-steroidal anti-inflammatory drugs have bacteriostatic and bactericidal activity against *Helicobacter pylori*. *Journal of gastroenterology and hepatology*. 2006 Sep;21(9):1388-93.
70. Reller LB, Weinstein M, Jorgensen JH, Ferraro MJ. Antimicrobial susceptibility testing: a review of general principles and contemporary practices. *Clinical infectious diseases*. 2009 Dec 1;49(11):1749-55.
71. Fouad AF, Nosrat A. Pulp regeneration in previously infected root canal space. *Endodontic Topics*. 2013 Mar;28(1):24-37.
72. Kim S, Kratchman S. Modern endodontic surgery concepts and practice: a review. *Journal of endodontics*. 2006 Jul 1;32(7):601-23
73. Schäfer E. Irrigation of the root canal. *Endodontic Practice Today*. 2007 Mar 1;1(1).
74. Spratt DA, Pratten J, Wilson M, Gulabivala K. An in vitro evaluation of the antimicrobial efficacy of irrigants on biofilms of root canal isolates. *International Endodontic Journal*. 2001 Jun;34(4):300-7.
75. Mohammaddi Z. Systemic, prophylactic and local application of antibiotics in endodontics: an update. *Int Dent J*. 2009;59:175-89.
76. Violich DR, Chandler NP. The smear layer in endodontics—a review. *International endodontic journal*. 2010 Jan;43(1):2-15.

77. Mahendra M, Agrawal N, Munaga S, Tyagi S. Antimicrobial activity of different biological extracts as intracanal medicament against *Enterococcus faecalis*: An in vitro study. *Endodontology*. 2016 Jul 1;28(2):166.
78. BystroByström A, Sunqvist G. The antibacterial action of sodium hypochlorite and EDTA in 60 cases of endodontic therapy. *International endodontic journal*. 1985 Jan;18(1):35-40.
79. Haapasalo M, Shen Y, Qian W, Gao Y. Irrigation in endodontics. *Dental Clinics*. 2010 Apr 1;54(2):291-312.
80. Sathorn C, Parashos P, Messer H. Antibacterial efficacy of calcium hydroxide intracanal dressing: a systematic review and meta-analysis. *International endodontic journal*. 2007 Jan;40(1):2-10.
81. Haapasalo HK, Sirén EK, Waltimo TM, Orstavik D, Haapasalo MP. Inactivation of local root canal medicaments by dentine: an in vitro study. *International endodontic journal*. 2000 Mar;33(2):126-31.
82. Weston CH, Barfield RD, Ruby JD, Litaker MS, McNeal SF, Eleazer PD. Comparison of preparation design and material thickness on microbial leakage through Cavit using a tooth model system. *Oral Surgery, Oral Medicine, Oral Pathology, Oral Radiology, and Endodontology*. 2008 Apr 1;105(4):530-5.
83. Sjögren U, Figdor D, Spångberg L, Sundqvist G. The antimicrobial effect of calcium hydroxide as a short-term intracanal dressing. *International endodontic journal*. 1991 May;24(3):119-25.
84. Deonizio MD, Sydney GB, Batista A, Estrela C. Root canal filling with calcium hydroxide paste using Lentulo spiral at different speeds. *Dental Press Endod*. 2011 Apr;1(1):58-63.





85. Chen G, Chang YC. Effects of liquid-and paste-type EDTA on smear-layer removal during rotary root-canal instrumentation. *Journal of Dental Sciences*. 2011 Mar 1;6(1):41-7.
86. Sungur DD, Aksel H, Purali N. Effect of a low surface tension vehicle on the dentinal tubule penetration of calcium hydroxide and triple antibiotic paste. *Journal of endodontics*. 2017 Mar 1;43(3):452-5.
87. Alam T, Nakazawa F, Nakajo K, Uematsu H, Hoshino E. Susceptibility of *Enterococcus faecalis* to a Combination of Antibacterial Drugs (3Mix) in vitro. *Journal of oral biosciences*. 2005;47(4):315-20.
88. SiqueiraJr JF, Lopes HP, de Uzeda M. Recontamination of coronally unsealed root canals medicated with camphorated paramonochlorophenol or calcium hydroxide pastes after saliva challenge. *Journal of endodontics*. 1998 Jan 1;24(1):11-4.
89. SiqueiraJr JF, Lopes HP, de Uzeda M. Recontamination of coronally unsealed root canals medicated with camphorated paramonochlorophenol or calcium hydroxide pastes after saliva challenge. *Journal of endodontics*. 1998 Jan 1;24(1):11-4.
90. Siqueira JF, de Uzeda M. Disinfection by calcium hydroxide pastes of dentinal tubules infected with two obligate and one facultative anaerobic bacteria. *Journal of Endodontics*. 1996 Dec 1;22(12):674-6.
91. Weiger R, De Lucena J, Decker HE, Löst C. Vitality status of microorganisms in infected human root dentine. *International endodontic journal*. 2002 Feb;35(2):166-71.

92. Weiger R, De Lucena J, Decker HE, Löst C. Vitality status of microorganisms in infected human root dentine. *International endodontic journal*. 2002 Feb;35(2):166-71.
93. Sedgley CM, Molander A, Flannagan SE, Nagel AC, Appelbe OK, Clewell DB, Dahlén G. Virulence, phenotype and genotype characteristics of endodontic *Enterococcus* spp. *Oral microbiology and immunology*. 2005 Feb;20(1):10-9.
94. Sedgley CM, Molander A, Flannagan SE, Nagel AC, Appelbe OK, Clewell DB, Dahlén G. Virulence, phenotype and genotype characteristics of endodontic *Enterococcus* spp. *Oral microbiology and immunology*. 2005 Feb;20(1):10-9.
95. Weiger R, De Lucena J, Decker HE, Löst C. Vitality status of microorganisms in infected human root dentine. *International endodontic journal*. 2002 Feb;35(2):166-71
96. Blanscet ML, Tordik PA, Goodell GG. An agar diffusion comparison of the antimicrobial effect of calcium hydroxide at five different concentrations with three different vehicles. *Journal of endodontics*. 2008 Oct 1;34(10):1246-8.
97. Riordan JT, Dupre JM, Cantore-Maty SA, Kumar-Singh A, Song Y, Zaman S, Horan S, Helal NS, Nagarajan V, Elasri MO, Wilkinson BJ. Alterations in the transcriptome and antibiotic susceptibility of *Staphylococcus aureus* grown in the presence of diclofenac. *Annals of clinical microbiology and antimicrobials*. 2011 Dec;10(1):1-1.
98. Wishart DS, Knox C, Guo AC, Cheng D, Shrivastava S, Tzur D, Gautam B, Hassanali M. DrugBank: a knowledgebase for drugs, drug actions and drug targets. *Nucleic acids research*. 2008 Jan 1;36(suppl\_1):D901-6.

99. Mazumdar K, Dastidar SG, Park JH, Dutta NK. The anti-inflammatory non-antibiotic helper compound diclofenac: an antibacterial drug target. *European journal of clinical microbiology & infectious diseases*. 2009 Aug 1;28(8):881.
100. Ulusoy S, Bosgelmez-Tinaz G. Nonsteroidal anti-inflammatory drugs reduce the production of quorum sensing regulated virulence factors and swarming motility in human pathogen *Pseudomonas aeruginosa*. *Drug research*. 2013 Aug;63(08):409-13.
101. Metri M, Hegde S, Bhandi S. Effect of pretreatment diclofenac sodium on postendodontic pain: A randomised controlled trial. *Journal of Conservative Dentistry: JCD*. 2016 Jan;19(1):7.
102. Sharma NK, Kindelan JD, Hutchinson D, Lancaster L. A study to compare ibuprofen effervescent granules with ibuprofen tablets in the treatment of acute dental pain. *Primary Dental Care: Journal of the Faculty of General Dental Practitioners (UK)*. 1994 Sep 1;1(1):5-8.



## ANNEXURE – I

## ETHICAL CLEARANCE CERTIFICATE

	<b>Research and Ethics Committee</b> <b>KLE V K INSTITUTE OF DENTAL SCIENCES</b> <b>KLE University</b>	
Accredited 'A' Grade by NAAC      Placed in Category 'A' by MHRD (Govt)		
Nehru Nagar, Belagavi - 590 010, Karnataka State		
☎: 0831-2470362 FAX: 0831-2470840	Web: <a href="http://www.kledental-bgm.edu.in">http://www.kledental-bgm.edu.in</a> E-mail: <a href="mailto:principal@kledental-bgm.edu.in">principal@kledental-bgm.edu.in</a>	
		SI. No. : 1221
<div style="border: 1px solid black; padding: 5px; display: inline-block;"><b>CERTIFICATE</b></div>		
<i>This is to Certify that the synopsis titled</i>		
<p><i>Comparison of antibacterial efficacy of calcium hydroxide, diclofenac with calcium hydroxide and ibuprofen with calcium hydroxide against enterococcus faecalis in an endodontic model - an in vitro study.</i></p>		
		<i>Submitted by</i>
<i>Dr. Deepti Ancy Chacko</i>		<i>P. G. Student /</i>
<i>Staff, Guided by Dr. Neha Shaded</i>		<i>from Department of</i>
<p><i>Conservative dentistry &amp; endodontics has been critically evaluated by committee members and granted ethical clearance to conduct the above mentioned study</i></p>		
Date : 24/06/2019		
 <b>Member Secretary</b> Research and Ethical Committee KLEVK Institute of Dental Sciences Belagavi	 <b>Chairman</b> Research and Ethical Committee KLEVK Institute of Dental Sciences Belagavi Chairman Research and Ethical Committee KLE V K Institute of Dental Sciences Belagavi	

## ANNEXURE – II

## BIOSTATISTIC CLEARANCE CERTIFICATE

	<b>KLE V.K. Institute of Dental Sciences</b>	
(A Constituent unit of KLE Academy of Higher Education & Research Deemed-to-be-University u/s 3 of the UGC Act, 1956) Nehru Nagar, Belagavi-590 010 INDIA		
Re-Accredited 'A' grade by NAAC (2 <sup>nd</sup> Cycle) & Placed in Category 'A' by MHRD (GoI)		
☎ 0831-2470362 FAX: 0831-2470640	Web: <a href="http://www.kledental-bgm.edu.in">http://www.kledental-bgm.edu.in</a> E-mail: <a href="mailto:principal@kledental-bgm.edu.in">principal@kledental-bgm.edu.in</a>	

*Biostatistics Clearance Certificate*


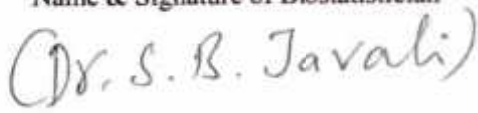
  

This is to certify that the Biostatistics aspect of the Dissertation / Research work of DR. DEEPTI ANCY CHACKO.....

entitled COMPARISON OF ANTI-BACTERIAL EFFICACY OF CALCIUM HYDROXIDE, DICLOFENAC WITH CALCIUM HYDROXIDE & IBUPROFEN WITH CALCIUM HYDROXIDE AGAINST E. FAECALIS- IN AN ENDODONTIC MODEL – AN IN VITRO STUDY



has been done under my guidance and considered satisfactory.

Place : Belagavi	 Name & Signature of Biostatistician
Date : 22/9/21	 (Dr. S. B. Javali)

## ANNEXURE – III

## PLAGIARISM CHECK CERTIFICATE

<b>Scientific Correspondence and Review Committee</b>	
<b>KLE VK Institute of Dental Sciences</b>	
A Constituent Unit of KLE Academy of Higher Education and Research (Deemed-to-be-University u/s 3 of the UGC Act, 1956) Nehru Nagar, Belagavi - 590 010, Karnataka State	
Accredited 'A' Grade by NAAC (2nd Cycle)	Placed in Category 'A' by MHRD (Govt)
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Date : 28 / 9 / 2020	Serial No. : 046
<b>PLAGIARISM CHECK REPORT</b>	
Name of the Applicant : Dr. Deepthi Anny Chacko	
UG / PG / Ph.D / Staff : Post graduate	
Batch & Year : 2018 - 2021	
Department : Conservative dentistry and Endodontics	
The soft copy of Research Work / Manuscript by <u>Dr. Deepthi Chacko</u> entitled "Comparison of antibacterial efficacy of Calcium hydroxide, "diclofenac" with Calcium hydroxide, ibuprofen with Calcium hydroxide against enterococcus faecalis in an "endodontic model an in vivo study" under the guidance of ..... has been submitted for Anti-Plagiarism check to the Scientific Correspondence & Review Committee of KLE VK Institute of Dental Sciences using "Turn-it-in" software.	
The scan has been carried out and the scanned output reveals a Similarity Index of .....3.....%, which is <b>within / not within</b> the acceptable limits of 10% as per the UGC guidelines.	
 <b>Member Secretary</b> Scientific Correspondence and Review Committee KLEVK Institute of Dental Sciences KAHER-Belagavi	 <b>Chairman</b> Scientific Correspondence and Review Committee KLEVK Institute of Dental Sciences KAHER - Belagavi